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Quantitative meta-analysis reveals no association between mercury contamination and body condition in birds

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ABSTRACT

Mercury contamination is a major threat to the global environment, and is still increasing in some regions despite international regulations. The methylated form of mercury is hazardous to biota, yet its sublethal effects are difficult to detect in wildlife. Body condition can vary in response to stressors, but previous studies have shown mixed effects of mercury on body condition in wildlife. Using birds as study organisms, we provide the first quantitative synthesis of the effect of mercury on body condition in animals. In addition, we explored the influence of intrinsic, extrinsic and methodological factors potentially explaining cross-study heterogeneity in results. We considered experimental and correlative studies carried out in adult birds and chicks, and mercury exposure inferred from blood and feathers. Most experimental investigations (90%) showed a significant relationship between mercury concentrations and body condition. Experimental exposure to mercury disrupted nutrient (fat) metabolism, metabolic rates, and food intake, resulting in either positive

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or negative associations with body condition. Correlative studies also showed either positive or negative associations, of which only 14% were statistically significant. Therefore, the overall effect of mercury concentrations on body condition was null in both experimental (estimate \pm SE = 0.262 \pm 0.309, 20 effect sizes, five species) and correlative studies (-0.011 \pm 0.020, 315 effect sizes, 145 species). The single and interactive effects of age class and tissue type were accounted for in meta-analytic models of the correlative data set, since chicks and adults, as well as blood and feathers, are known to behave differently in terms of mercury accumulation and health effects. Of the 15 moderators tested, only wintering status explained cross-study heterogeneity in the correlative data set: free-ranging wintering birds were more likely to show a negative association between mercury and body condition. However, wintering effect sizes were limited to passerines, further studies should thus confirm this trend in other taxa. Collectively, our results suggest that (*i*) effects of mercury on body condition are weak and mostly detectable under controlled conditions, and (*ii*) body condition indices are unreliable indicators of mercury sublethal effects in the wild. Food availability, feeding rates and other sources of variation that are challenging to quantify likely confound the association between mercury and body condition in *natura*. Future studies could explore the metabolic effects of mercury further using designs that allow for the estimation and/or manipulation of food intake in both wild and captive birds, especially in under-represented life-history stages such as migration and overwintering.

Key words: blood, body mass, fat, feather, feeding, energetics, habitat, metabolism, species

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I INTRODUCTION

Chemical pollution is a major anthropogenic modification of the global environment, and a fundamental characteristic of the Anthropocene (Lewis & Maslin, 2015). Humans are responsible for the synthesis and release of a plethora of chemical contaminants for agricultural uses [e.g. organochlorine and organophosphate pesticides (Jones & De Voogt, 1999; Sánchez-Santed, Colomina & Herrero Hernández, 2016)], and industrial or every-day life applications [e.g. metallic trace elements, perfluoroalkyl substances, chlorinated paraffins (Walker *et al.*, 2012; Sunderland *et al.*, 2019; Vorkamp *et al.*, 2019)]. Among these contaminants, and despite being a natural element, mercury (Hg) is particularly hazardous to humans and wildlife because it has no biological function and is highly toxic even at low concentrations (Walker *et al.*, 2012; UN Environment, 2019). In its inorganic form, Hg has an atmospheric lifetime of 0.5–1 year and can be transported over vast spatial scales (Obrist *et al.*, 2018; UN Environment, 2019). Once deposited on the Earth's surface, Hg can undergo complex, microbially mediated processes and be converted into methylmercury (MeHg), which is assimilated and accumulated by

living organisms, and biomagnifies in food webs (Atwell, Hobson & Welch, 1998; Evers *et al.*, 2016; Eagles-Smith *et al.*, 2018).

Anthropogenic Hg emissions have declined in North America and Europe over the past two decades (UN Environment, 2019), but are still increasing in East Asia and in the Southern Hemisphere (Obrist et al., 2018). In addition, concentrations measured in wildlife are still increasing in several regions of the Northern (Braune et al., 2014; Wang et al., 2019) and Southern Hemispheres (Mills et al., 2020; Seco et al., 2020). The Minamata Convention, an international treaty that came into force in 2017 (http://www. mercuryconvention.org), adopted a global strategy to reduce Hg emissions and protect human and environmental health. While these global restrictions may limit increases in future Hg emissions linked to economic growth, legacy Hg emissions will continue to affect the Hg cycle for decades to centuries (Eagles-Smith et al., 2018). Monitoring Hg concentrations and effects in biota and the environment is thus a priority to oversee the effectiveness of the Minamata Convention (Evers et al., 2016).

Adverse effects of Hg in humans and wildlife include neurological, endocrine, and immune disruption with consequences on development, neurocognitive function, and reproduction (Tchounwou et al., 2003; Heinz et al., 2009; Tan, Meiller & Mahaffey, 2009; Tartu et al., 2013; Goutte et al., 2014; Eagles-Smith et al., 2018; Evers, 2018). Several intrinsic and extrinsic factors drive variation in Hg contamination and kinetics in wild organisms. Feeding ecology is a key explanatory factor of among- and within-species variation in tissue Hg concentrations (Anderson et al., 2009; Carravieri et al., 2014b, 2021; Polito et al., 2016; Ma et al., 2021) but other traits such as sex and age can also modulate this variation (Eagles-Smith et al., 2009; Robinson, Lajeunesse & Forbes, 2012; Jackson et al., 2015; Chételat et al., 2020). In addition, life-history traits such as breeding or migration strategies can influence diet, feeding rate, energy storage and expenditure, thus driving variation in Hg burdens (Seewagen, Cristol & Gerson, 2016; Ackerman, Hartman & Herzog, 2019; Adams et al., 2020a). All these intrinsic and extrinsic factors thus have the potential to modulate Hg toxicity. However, identifying sublethal effects of Hg in the field can be challenging, due to the potentially confounding influence of concurring environmental stressors (Marcogliese & Pietrock, 2011; Marteinson, Marcogliese & Verreault, 2017; Bårdsen, Hanssen & Bustnes, 2018). Studies have shown species-specific sensitivity to Hg toxicity [e.g. embryotoxicity in birds (Heinz et al., 2009); neuroreceptor inhibition in mammals (Basu et al., 2005)], but the key phylogenetic and life-history traits or environmental factors that could explain these differences have yet to be identified clearly. Meta-analytical approaches quantifying the link between Hg and health endpoints in a large number of taxa could be effective in identifying the interactive factors that limit our capacity to detect significant effects of Hg on wildlife health.

Here, we performed a meta-analysis investigating the effect of Hg contamination on body condition using information extracted from 47 studies on 147 species of birds. Body condition indices are available in a large number of studies and species, and they can be calculated from morphometric measures that are used routinely in avian investigations. Albeit widely used, body condition indices may not be reliable indicators of health status (Fischer, Taborsky & Dieckmann, 2009; Schultner et al., 2013). Here, we consider body condition as an integrative measure of fat and lean mass (Peig & Green, 2009; Labocha & Hayes, 2012) that can be affected linearly or non-linearly by stressors (Pravosudov & Grubb, 1997; Schultner et al., 2013), for example via changes in behavfeeding performance) and/or physiology iour (e.g. (e.g. disruption of nutrient metabolism and energy use). We chose birds as the study taxon because they have served widely as early sentinels of negative effects of Hg on wildlife and ecosystem health (Wolfe, Schwarzbach & Sulaiman, 1998; Whitney & Cristol, 2018). Birds are ubiquitous, being found in a large variety of terrestrial, freshwater and marine habitats from polar regions to the tropics. In addition, they belong to different dietary guilds, from herbivores to omnivores, are relatively accessible compared to other groups of vertebrates, and are thus extensively studied (Konishi et al., 1989). Our aim was twofold: (i) to test for a systematic trend in the effect of Hg on body condition in birds; and (ii) to identify moderators of this relationship among intrinsic (e.g. species, age class, sex) and extrinsic factors (habitat type, dietary guild), as well as methodological aspects (tissue used to measure Hg concentrations, correlative or experimental approach). As Hg can affect bird behaviour [e.g. reduced ability to forage and/or compete for food (e.g. Evers et al., 2008)], metabolic rates (Gerson, Cristol & Seewagen, 2019), and physiological pathways involved in the stress response (Wada et al., 2009; Franceschini et al., 2017), which can have a range of contrasting consequences for physical condition, we expected either a positive or negative relationship between Hg concentrations and body condition. Given Hg biomagnification and naturally high bioavailability in aquatic environments (Atwell et al., 1998; Fitzgerald, Lamborg & Hammerschmidt, 2007), piscivorous species can be at greater risk of Hg exposure (Scheuhammer et al., 2007), and are thought to have developed a better tolerance to toxicity over evolutionary timescales [e.g. through more efficient detoxification mechanisms (Robinson et al., 2011; Manceau et al., 2021)]. Piscivorous species might therefore show a smaller effect size of the Hg-body condition association.

II MATERIALS AND METHODS

(1) Search and inclusion criteria

Our literature search was conducted in ISI *Web of Science* (latest search 22/01/2021) across all years, using the search terms "mercury", "bird" and "body condition". Since body condition is often calculated in studies of the effect of Hg on physiological and fitness endpoints without being the primary objective, we also searched the literature using the terms "mercury", "bird" and one of the following key words:



Fig. 1. PRISMA© diagram describing the different phases of the systemic review on the association between Hg contamination and body condition, where N is the number of studies. *Some data sets were published in more than one scientific article, see Table 1.

"effect", "health", "telomere", "oxidative stress", "hormone", "corticosterone", "testosterone", "thyroid", "immunity", "parasite", "DNA damage", "energy", "hatching/ fledging/breeding success", "hatch date", "clutch size", "foraging behaviour", "survival", "growth" and "body mass". The reference lists of literature reviews found in this way were also checked to expand the database (Fig. 1). We contacted the authors of studies to obtain further details when (i) statistical information was missing, and/or (ii) the effect size was calculated across several species. Incomplete statistical information and a lack of a response by contacted authors meant that some relevant studies had to be discarded (see online Supporting Information, Table S1). The literature search resulted in the selection of 47 studies spanning publication years 2000 to 2020 (Table 1). These covered data mainly on passerines and seabirds with a few waders and raptors (Table 1). In addition, we included nine studies from the authors' unpublished work in order to reduce the bias towards passerine birds. Overall, our meta-analysis included 147 species of birds, which were mainly passerines and seabirds. Inclusion criteria are detailed below.

(a) Body condition indices

Previous studies attempting to determine which body condition index better represents fat and/or lean mass have produced mixed results in both mammals and birds (e.g. Labocha, Schutz & Hayes, 2014; Kraft *et al.*, 2019). Here, variables initially retained as pertinent indices of body condition were: size-corrected body mass [e.g. scaled mass index (SMI), structural size to body mass ratio, residuals of an ordinary least-squares regression of body mass against a linear morphometric measure of size], body mass, organ-tobody ratio, fat scores, pectoral muscle thickness, and organ masses. However, only eight effect sizes from four studies were estimated from indices other than size-corrected body mass or body mass (Table S1), preventing an accurate estimation of their potential influence on the Hg-body condition relationship. Therefore, these effect sizes were not included in meta-analytic models. The majority of published articles from which effect sizes were extracted used SMI (Peig & Green, 2009) as body condition index (see Table S2 for the frequency of all condition indices). Therefore, to minimise data heterogeneity, we also calculated SMI as the body condition index when we had access to raw data. The SMI has several advantages over other indices because it is not size dependent and can be used readily to compare across populations (Labocha *et al.*, 2014). The SMI adjusts the mass of the individuals to the mass they would have if all individuals had the same body size, using the following equation:

$\dot{M}_i = M_i L_0^{b \text{SMA}} / L_i$

where M_i and L_i are the body mass and the body length measure of individual i, respectively; the exponent b_{SMA} is estimated by the standardised major axis (SMA) regression of log body mass on log body length; L_0 is an arbitrary value of body length, and M_i is the predicted body mass for individual *i* when the body length is standardised to L_0 . We calculated L_0 as the arithmetic mean of the body length variable chosen for each study. As different measures of body length (e.g. tarsus, bill, and wing length) can scale differently with body mass depending on species, we used the body size measure selected by the authors to calculate L_0 and thus SMI. When no preference was communicated, and several body length measures were available, we used tarsus or bill length rather than wing length, which is difficult to measure reproducibly and can be a poor indicator of structural body size (Jenni & Winkler, 1989). The choice between tarsus or bill length was made by taking the measure that correlated better with body mass on a log basis, as this is likely to be the best one explaining the fraction of mass associated with structural size (Peig & Green, 2009).

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Reference*	Year	Location	Species	ES	Tissue	Age class	Condition type
Correlative studies Ackerman <i>et al.</i> (2019)	2012, 2013	Central Valley of California	40 passerine species	78	Blood &	Adult	Body
Adams et al. (2020a)	2013-2017	Multiple sites in New York State, USA	54 passerine species	54	Feather Blood	Adult	condition Body
Adams et al. (2020b)	2009–2012	Bill Baggs Cape Florida State Park	8 passerine species	12	Blood	Adult	condition Body
Amélineau <i>et al.</i> (2019)	2004 - 2015	East Greenland	Alle alle	-	Feather	Adult	condition Body
Unpublished O.G., G.Y.*	2018, 2019	North-East Greenland	Pagophila eburnea	7	Blood	Adult	$\operatorname{condition}_{\operatorname{Body}}$
Unpublished E.G.*	2017, 2018	Talan Island, Sea of Okhotsk, Russia	2 seabird species	5	Feather	Adult	condition Body
Unpublished H.S.*	2015 - 2018	Bjornoya Island, Barents Sea, Norway	4 seabird species	7	Blood &	Adult	condition Body
Unpublished A.P.W., A.K.*	2015, 2016	Saint Lawrence Island, Bering Sea	6 seabird species	11	Feather Blood & Ft	Adult	condition Body
Carravieri <i>et al.</i> (2018) Carravieri <i>et al.</i> (2020a)	2011-2014 2018	Dronning Maud Land, Antarctica Isle of May, Firth of Forth, Scotland	Thalassoica antarctica Phalacrocorax aristotelis	2 2	reamer Blood Blood	Adult Adult	condition Body mass Body
Eckbo et al. (2019)	2015	Svalbard, Norwegian Arctic	Cepphus grylle mandtii	-	Blood	Adult	condition Body
Unpublished J.F.*	2019	Tromelin & Ile dy Lys, Indian Ocean	3 seabird species	9	Blood	Adult	$\operatorname{condition}_{\operatorname{max}}$
Gurney <i>a al.</i> (2014) Hargreaves <i>a al.</i> (2010, 2011)	2002, 2003 2009	Redberry Lake, Canada Southampton Island, Nunavut	<i>Melanitta fusca</i> 6 wader species	1	Blood Blood	Adult Adult	condition Body mass Body
Unpublished K.L.* Koiadinovic <i>et al.</i> (2007a)	2017 2004	Karrak Lake, Nunavut Multinle sites. Western Indian Ocean	2 wader species Omehotrion fueratus	сı –	Blood Feather	Adult Adult	condition Body mass Body mass
Provencher $et al.$ (2016a)	2013, 2014	Mittivik Is, Northern Hudson Bay,	Somateria mollissima		Blood	Adult	Body condition
Rowse et al. (2014)	2011, 2012	Upper Scioto River, Ohio, USA	Empidonax virescens	Ч	Blood	Adult	Body
Scheuhammer et al. (2016)	2008	Manitoba & Saskatchewan, Canada	Gavia immer	4	Blood & Feather	Adult	condition Body mass
Seewagen (2013) Unpublished C.L.S.* Soldatini <i>et al.</i> (2020)	2008, 2009 2007, 2008 2017	New York, USA New York, USA Natividad Island, Mexico	Parkesia noveboracensis Hylocichla mustelina Puffinus opisthomelas		Blood Blood Blood	Adult Adult Adult	Body mass Body mass Body
Tartu <i>et al.</i> (2013)	2008, 2011	Svalbard, Norwegian Arctic	Rissa tridactyla	7	Blood	Adult	condition Body
Tartu $et al. (2014)$	2008	Terre Adéie, Antarctica	Pagodroma nivea	5	Blood	Adult	Body Boddition
Tartu <i>et al.</i> (2015)	2010	Terre Adélie, Antarctica	Pagodroma nivea	2	Blood	Adult	COLICITOR

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Reference*	Year	Location	Species	ES	Tissue	Age class	Condition type
		-	-				Body condition
Tartu et al. (2016)	2012, 2013	Svalbard, Norwegian Arctic	Rıssa trıdactyla	51	Blood	Adult	Body condition
Albertos et al. (2020) Ackerman et al. (2012)	2005-2020 2006-2010	Alicante, Spain South San Francisco Bay, USA	2 seabird species Rallus longirostris obsoletus	2 2	Feather Blood &	Adult Adult	Body mass Body
Wavland <i>et al.</i> (2001–2002–2003.	1997, 1998	East Bay Mieratory Bird Sanctuary.	Somateria mollis sima	_	Feather Blood	Adult	condition Body
2005)		Nunavut					condition
Lerma et al. (2016)	2011, 2012	Bahia Santa Maria, Sinaloa, Mexico	Sula nebouxii	ŝ	Blood	Adult &	Body
Unpublished P.B., A.C., O.C., Y.C.*	2012	French Austral Territories, Southern	15 seabird species	52	Blood &	Cnick Adult &	conatuon Body
Sebastiano et al. (2017)	2013	Ocean Grand Connetable. French Guvana	6 seabird species	11	Feather Blood	Chick Adult &	condition Bodv
						Chick	condition
Weech $et al.$ (2006)	2000–2002	lakes, British Columbia	Haliaeetus leucocephalus	5	Blood	Adult & Chicb	Body mass
Clarkson et al. (2012)	2009, 2010	Virginia and New York, USA	1 seabird, 1 wader	5	Feather	Chick	Body mass
Costantini et al. (2020)	2016-2018	Linosa. Italy	species Calonectris diomedea	2	Blood	Chick	Body mass
Carravieri et al. $(2020b)$	2012	French Austral Territories, Southern	13 seabird species	13	Blood	Chick	Body
Unpublished P.B., J.F.*	2015-2017	Ocean French Coast, English Channel	4 seabird species	œ	Blood &	Chick	condition Body
Carravieri <i>et al.</i> (2017)	9019	French Austral Territories Southern	9 seahird species	с с	Feather Blood	Chick	condition Bodv
	1	Ocean		5	0000		condition
Herring et al. (2014)	2006, 2007	Everglades, Florida	2 wader species	5	Blood	Chick	Body
Kojadinovic et al. (2007b)	2002–2004	Reunion Island, Western Indian Ocean	2 seabird species	5	Feather	Chick	Body
Ortiz-Santaliestra et al. (2015)	2010, 2011	Castilla and Leon, Andalusia, Spain	Aquila fasciata	4	Blood	Chick	condition Body
Santos et al. (2017)	2012, 2013	Belgium	Larus fuscus	1	Feather	Chick	condition Body
Santos et al. (2020)	2015	Ostend, Belgium	Larus fuscus	-	Feather	Chick	condition Body mass
Experimental studies Seewagen <i>et al.</i> (2016); Gerson <i>et al.</i> (9010)	2015	Laboratory	Taeniopygia guttata	4	Blood	Adult	Body mass
(2013) Kobiela <i>et al.</i> (2015)	2014	Laboratory	Taeniopygia guttata	5	Blood	Adult	Body
Seewagen et al. (2019)	2016-2017	Long Point, Lake Erie, Canada	Setophaga coronata	1	Blood	Adult	Body
Ma et al. (2018)	2014-2017	Long Point, Lake Erie, Canada	Setophaga coronata	4	Blood	Adult	Body mass
							(Continues)

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Table 1. (Cont.)

Reference*	Year	Location	Species	ES	Tissue	Age class	Condition type
Kenow et al. (2003)	1999, 2000	Laboratory	Gavia immer	-	Blood	Chick	Body
Fallacara <i>et al.</i> (2011)	2005	Laboratory	Falco sparverius	7	Blood	Chick	Body
Spalding et al. (2000)	1996	Everglades, Florida	Ardea alba	5	Blood	Chick	Body
Yu et al. (2016)	2015	Laboratory	Taeniopygia guttata	4	Blood	Chick	condutor Body conditior

(b) Hg chemical form

The form of Hg in avian diets may vary depending on prey types. High-trophic-level prey, such as fish and squid, mainly contain Hg as MeHg, whereas invertebrates may have higher proportions of inorganic Hg (Bloom, 1992; Mason, Laporte & Andres, 2000; Bustamante *et al.*, 2006; Seco *et al.*, 2020). All included studies measured total Hg, which is an accurate proxy of MeHg in tissues such as blood and feathers (e.g. Bond & Diamond, 2009; Renedo *et al.*, 2021).

(c) Exposure pathway and measure

We included experimental studies where adults and chicks were exposed to Hg via dietary exposure as MeHg, as well as MeHg egg injections. We considered studies where Hg contamination was inferred from concentrations [in parts per million (ppm) or µg per unit wet or dry mass) measured in whole blood, red blood cells, and feathers, in adult birds and chicks. We excluded effect sizes obtained from other tissues given their heterogeneity and small number (Table S1). In experimental studies all Hg concentrations were reported on a wet mass basis. By contrast, the majority of correlative studies reported Hg concentrations on a dry mass basis. In order to provide homogenous Hg concentration estimates, correlative studies presenting results on a wet mass basis were converted to dry mass. The latter calculation was based on a moisture content of 65% in red blood cells (P. Bustamante & O. Chastel, unpublished data), and 77% in whole blood [mean of moisture values measured in Eagles-Smith et al. (2008), Ackerman, Hartman & Herzog (2017) and Ackerman et al. (2019)]. Blood is a better representative of Hg body burden than are feathers in birds, and should be preferred in toxicity risk estimations (Fuchsman et al., 2017; Chételat et al., 2020). In addition, Hg temporal integration into feathers can be highly variable depending on species, feather type, moult strategy and moult stage at sampling (Carravieri et al., 2014a; Albert et al., 2019; Peterson et al., 2019). However, in some species, feather Hg concentrations correlate well with concentrations in internal tissues, including blood, and can thus also be useful representatives of Hg body burdens (Ackerman et al., 2012, 2019; Fort et al., 2014). Therefore, we decided to include studies reporting associations between feather Hg concentrations and body condition in our meta-analysis, and test tissue type (blood or feather) as a moderator. Body feathers are preferentially sampled for ethical and practical reasons, and, to avoid further heterogeneity in the data set, we excluded the few effect sizes obtained from other feather types (Table S1).

(d) Other criteria

Studies comparing body condition indices between populations at polluted sites where pollution was not clearly Hg-related were also excluded, as well as studies on dead, emaciated individuals (Table S1), where body condition could be biased. Effect sizes obtained from less than four individual birds were discarded.

(2) Moderators included and categorisation

Our meta-analysis included a maximum of 15 moderators: (i) bird type (passerine, raptor, seabird, wader); (ii) age class (chick, adult); (iii) whether the effect size had been corrected for sex; (iv) whether the effect size had been corrected for other factors that were specific to the study (e.g. individual random factor, sampling date, season); (v) dietary guild (carnivore, herbivore, invertivore, omnivore); (vi) habitat type (freshwater, marine, terrestrial); (vii) geographical zone (polar/subpolar, temperate, tropical/subtropical); whether the sampled population was (viii) wintering (ves/no). (ix) migrating (ves/no) or (x) breeding (ves/no); (xi) tissue type used for Hg quantification (blood, feather); (xii) body condition index (body mass, size-corrected body mass); (xiii) species-specific basal metabolic rate (BMR); (xiv) Hg concentration; (xv) ratio of species-specific maximum body mass to average body mass (hereafter BM ratio, which reflects the maximum body condition of the species; Vincze et al., 2019). All moderators, their modalities, the number of effect sizes, and the justification for including them to study the link between Hg concentration and body condition are reported in Table 2. Dietary guild was assigned based on results in the sampled population of each study when available, or from the Wilman et al. (2014) database. Species-specific BMR values were extracted from Ellis & Gabrielsen (2001), Møller (2009), Londoño et al. (2015) and McKechnie, Noakes & Smit (2015), but were not available for all species. Body mass information to calculate the BM ratio was extracted from the Dunning (2007) database, or from additional references (Glahn & McCoy, 1995; Shirihai et al., 2002; Kooyman et al., 2004; Helseth, Stervander & Waldenström, 2005; Tobón & Osorno, 2006; Kojadinovic et al., 2007a; Overton et al., 2009; Hancock, Kushlan & Kahl, 1992; Rising, 2010; García, Moreno-Opo & Tintó, 2013; Maccarone & Brzorad, 2016; studies included in the metaanalysis). For each study, we extracted the moderators, as well as sample and effect sizes as detailed below.

(3) Effect size extraction or calculation

Given that most studies were correlative, we chose Pearson's r as the effect size. Based on the information given in correlative studies, we calculated effect sizes using t-values, F-values, P-values, means and standard deviations, or correlation coefficients, following formulae given in Koricheva, Gurevitch & Mengersen (2013). For studies reporting the comparison of experimental groups, we calculated standardised mean differences (Cohen's d), which were then transformed to Pearson's r, following Koricheva et al. (2013). To adhere to normality assumptions, Pearson's r were then converted to Fisher's Zr following the equation in Lipsey & Wilson (2001). Sampling variances associated Z-scores were calculated as $(n-3)^{-1}$ following Koricheva *et al.* (2013). Positive effect sizes indicate a positive effect of Hg on body condition, while negative effect sizes denote a decrease in body condition with increasing Hg concentrations.

Table 2. List of moderators included in the meta-analysis

Moderator	Modalities (number of effect sizes) ^a	Expected influence on the Hg–body condition association
Bird type ^b	Passerine (145; 15), Raptor (6; 2), Seabird (150; 1), Wader (14; 2)	Physiological or life- history related differences in sensitivity to Hg
Age class	Adult (256; 11), Chick (59; 9)	Age-related sensitivity to Hg
Accounted for sex	Yes (18; 5), No (297; 15)	Sex-related sensitivity to Hg
Accounted for other variables	Yes (6; 11), No (309; 9)	Accounting for other confounding factors may modulate effect sizes
Dietary guild	Carnivore (114; 5), Herbivore (25; 10), Invertivore (143; 5), Omnivore (33; 0)	Potential exposure to different Hg chemical forms
Habitat type ^b	Freshwater (9; 1), Marine (153; 0), Terrestrial (153; 19)	Potential exposure to different Hg chemical forms
Zone ^b	Polar/subpolar (105; 0), Temperate (156; 18), Tropical/subtropical (54, 2)	Different energetic demands may change susceptibility to Hg
Wintering ^b Migrating Breeding ^b	Yes (16; 0), No (299; 20) Yes (41; 5), No (274; 15) Yes (197; 0), No (118; 20)	Physiological status, energetic demands and behaviour of different life-history stages may change susceptibility to Hg
Tissue ^b	Blood (213; 20), Feather (102; 0)	Tissue-specific temporal integration of Hg
Body condition index ^b	Body mass (19; 17), Size-corrected body mass (296; 3)	Index-associated influence on the Hg–body condition relationship
Covariate	Unit (number of	F
Hg concentration	$\mu g/g (315; 20) (dw and ww for correlative and experimental studies, respectively)$	Concentration- dependent susceptibility to Hg
BMR	kJ/day (132; 17)	Energetic needs may influence the susceptibility to Ha
BM ratio	No unit (315; 20)	Potential effect of fat load on body condition

BM, body mass; BMR, basal metabolic rate; dw, dry mass; ww, wet mass.

^bModerators not included in meta-analytic models of experimental studies given the limited sample size.

(4) Meta-analytic technique

In a preliminary step, we conducted a meta-analysis on the full data set including 335 effect sizes and a study type

^{ar}The first and second number are for correlative and experimental studies, respectively.

moderator (correlative or experimental) (results not shown). However, experimental studies appeared to be highly influential in the effect estimates, because of extreme values and high leverage. Therefore, we carried out separate metaanalyses for correlative and experimental data sets.

(a) Potential bias in effect size reporting

To test for the presence of potential bias in the results of our meta-analysis, we first explored funnel plots produced between effect sizes and their corresponding measure of precision (here standard error, SE) for signs of asymmetry. Second, we performed Egger's regression tests (Egger et al., 1997) using overall intercept-only models with the regtest function (metafor package, v. 2.4-0; Viechtbauer, 2010). Egger's test evaluates the relationship between effect sizes and measurements of study precision (Egger et al., 1997), and can reveal different types of bias, such as reporting bias or poor methodological quality (Sterne et al., 2011). For instance, studies reporting a significant effect may be more likely to be published than studies reporting no effect (Koricheva et al., 2013; but see Koricheva, 2003). Here, most effect sizes for correlative studies, and all effect sizes for experimental studies, were extracted from published resources (Fig. 1). Our meta-analysis also included unpublished resources for the correlative data set. We thus tested (i) potential bias in published and unpublished studies separately, as well as combined; (ii) whether the publication status (used as a moderator) of the effect size had an effect on the meta-analytical results.

(b) Random structure and overall effect of Hg on body condition

All meta-analytic models were performed using the *ma.mv* function in the *metafor* package (v. 2.4-0; Viechtbauer, 2010) in R (v. 4.0.5; R Core Team, 2021). We constructed multilevel meta-analytic linear mixed effect models, which facilitates the control of multiple sources of non-independence. Our data were affected by multiple types of non-independence (Noble et al., 2017), including non-independence of effect sizes, multiple effect sizes originating from the same studies, non-independence of observations of the same species, as well as non-independence of species (shared ancestry). We aimed to control for these dependencies by testing the effect of three random variables: individual effect size identity (individual effect size ID, unique per data row, necessary to estimate residual heterogeneity; Noble et al., 2017), study identity (study ID), and species identity (species ID). In the correlative data set, we also tested the influence of the phylogenetic variance-covariance matrix representing the phylogenetic history of the species. For the latter, we used a rooted ultrametric consensus tree that was inferred from the SumTrees Python library (Sukumaran & Holder, 2010), based on 1000 random trees obtained from birdtree.org (Jetz et al., 2012), using the Hackett backbone tree (Hackett et al., 2008). Phylogeny was not accounted for in the experimental data set, as including phylogenetic random effects with less than 15 species can lead to unreliable estimates (Bolker et al., 2009). To select the appropriate random structure, we constructed intercept-only meta-regression with all combinations of the three random variables, as well as the phylogenetic signal for correlative studies, using the maximum-likelihood (ML) method. We then compared the models using the Akaike Information Criterion (AIC) and chose the random structure of the model with the lowest AIC value, and lowest number of variables when AIC values were similar between models (parsimony criteria) (Table S3). Using the random structure of the best selected model, we tested the overall effect size of Hg contamination on body condition using restricted maximum likelihood approximation (REML). We also calculated the heterogeneity statistic (I_{total}^2) based on an intercept-only meta-analytic model (built using the rma function in *metafor*), without any random effects. I^2 represents the percentage heterogeneity in the effect sizes (0-100%) due to true heterogeneity rather than random sampling variance (Higgins & Thompson, 2002; Higgins *et al.*, 2003). I^2 was moderate to high (defined as $l^2 > 50\%$; Higgins *et al.*, 2003; see Section III), therefore we ran moderator analyses to explore which life-history, ecological or physiological parameters could explain the high heterogeneity observed in the effect of Hg contamination on body condition in birds.

(c) Moderator analysis

In order to study the influence of different moderators on the association between Hg contamination and body condition, we constructed multifactorial meta-analytic models using the MuMIn package (Bartón, 2020). Chicks and adults, as well as blood and feathers, are known to behave differently in terms of Hg accumulation and health effects (e.g. Whitney & Cristol, 2018), and could show different associations between Hg and body condition. Therefore, their single and interactive effects were always accounted for in models of the correlative data set, as follows: \mathcal{Z} score ~ age class + tissue type + age class*tissue type + Moderator (Table S4). Models were constructed with the selected random structure (see Sections II.4b and III), fitted using ML, and compared to the base model: \mathcal{Z} score ~ age class + tissue type + age class*tissue type. In preliminary steps the age class 'juveniles' was considered, but data were too scarce to enable inclusion in the final meta-analysis. The available data on body condition and Hg concentrations in blood or feathers of juveniles were pooled with those of adults. We considered that a moderator had a significant effect on the association between Hg and body condition when its addition to the base model decreased the AIC corrected for small sample size (AICc) by at least 2. Data on BMR were available only for a subset of observations. To test the effect of BMR on the association between Hg and body condition we thus constructed a separate set of models (Table S4). Using these models, we obtained parameter estimates for each predictor after refitting models with REML.

The experimental data set was too small for multifactorial statistical analysis (N = 20 effect sizes). Therefore, each

0.094 0.25 Standard error Standard e rror 0.189 0.5 0.283 0.75 0.378 2 3 -2 -1 0 1 2 -3 -2 -1 0 1 Observed outcome Observed outcome

0

Fig. 2. Funnel plot for correlative (left) and experimental (right) studies.

moderator was entered as a single predictor to an interceptonly multilevel meta-analytic model (fitted by REML) with the selected random structure (Table S5). Using these models, we obtained parameter estimates for each predictor and factor level. The overall significance of each predictor was assessed using an omnibus test (Viechtbauer, 2010).

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III RESULTS

(1) Bias in effect size reporting

For the correlative data set, Egger's tests indicated a tendency for biased reporting towards negative effect sizes among published studies (Egger's test: $\mathcal{N} = 224$, z = -1.932, P = 0.053), and a significant bias towards positive effect sizes among unpublished studies (Egger's test $\mathcal{N} = 91$, z = 2.148, P = 0.032; see Table S6 for funnel plots). By contrast, we found no statistical evidence of bias in the correlative database combining published and unpublished effect sizes (Egger's test: $\mathcal{N} = 315$, z = -0.707, P = 0.480). In addition, publication status had no effect on the Hg-body condition association (Table S6). Therefore, all further meta-analytic models were only run on the full correlative data set combining published and unpublished effect sizes.

We detected a tendency for biased reporting towards negative effect sizes in experimental studies (Fig. 2; Egger's test: $\mathcal{N} = 20, \ z = -1.894, \ P = 0.058$), but this bias was nonsignificant.

(2) Correlative studies

We used 315 population-specific effect sizes from 145 species to test the relationship between Hg concentration and body condition in correlative studies. These were carried out in wild populations, including mainly adult passerines and seabirds, from terrestrial and marine environments, in temperate and polar/subpolar regions (Table 2). The median number of individuals inspected per effect size was 16 (range: 4–1051). Comparison of intercept-only multilevel meta-analytic models indicated a significant increase in fit with the inclusion of individual effect size ID and species ID (Table S3), which were retained in the random structure of all subsequent models. Phylogenetic signal appeared to have little influence on the overall effect size and did not affect model fit. The overall effect of Hg contamination on body condition was nonsignificant (estimate \pm SE = -0.011 ± 0.020 , confidence interval, CI [-0.049; 0.027], while accounting for individual effect size ID and species ID). Effect size heterogeneity was moderate $(I^2 = 55\%)$; Cochran's Q test = 668, df = 314, P < 0.0001), indicating suitability for moderator analyses. Multifactorial moderator analyses revealed that wintering status had an influence on effect size estimates (AICc 2.6 points lower than the base model, Table S4): wintering birds were more likely to show a negative effect of Hg on body condition (Fig. 3). Other moderators had no clear effects on the Hg-body condition association (Table S4).

(3) Experimental studies

Experimental studies encompassed 20 effect sizes from five species (Ardea alba, Falco sparverius, Gavia immer, Setophaga coronata, Taeniopygia guttata). The median number of individuals inspected per effect size was 24 (range: 5–49). Only individual effect size ID was retained in the random structure of intercept-only multilevel meta-analytic models (Table S3). Hg contamination was not related to body condition (0.262 \pm 0.309, [-0.344; 0.867], while accounting for individual effect size ID), and effect size heterogeneity was very high ($I^2 = 99\%$; Cochran's Q test = 887, df = 19, P < 0.0001). In the experimental data set, no moderator predicted the Hg–body condition relationship (Table S5, Fig. 4, P > 0.100 in all Omnibus tests).

IV DISCUSSION

Our extensive meta-analysis showed no overall effect of Hg contamination on body condition across 147 free-



Explanatory	Sample size	Level	95% confidence interval	p-value	
Age class * Tissue + bird type	315	Passerine	[-0.082-0.033]	0.406	
		Raptor	[-0.284-0.252]	0.907	
		Seabird	[-0.035–0.103]	0.334	
		Wader	[-0.307-0.019]	0.084	
Age class * Tissue + habitat type	315	Freshwater	[0.001-0.380]	0.049	-
		Marine	[-0.057-0.078]	0.765	
		Terrestrial	[-0.087-0.024]	0.261	-8-
Age class * Tissue + zone	315	Polar/subpolar	[-0.024-0.130]	0.175	
		Temperate	[-0.088-0.024]	0.266	-8-
		Tropical/subtropical	[-0.119-0.060]	0.519	— —
Age class * Tissue + dietary guild	315	Carnivore	[-0.069-0.093]	0.773	
		Herbivore	[-0.251-0.021]	0.098	
		Invertivore	[-0.062-0.056]	0.922	
		Omnivore	[-0.128-0.111]	0.885	
Age class * Tissue + other var. acc.	315	No	[-0.054-0.042]	0.805	-8-
		Yes	[-0.264-0.102]	0.383	- _
Age class * Tissue + sex acc.	315	No	[-0.059-0.039]	0.698	
		Yes	[-0.123-0.107]	0.892	
Age class * Tissue + body cond. index	315	Body mass	[-0.247-0.054]	0.208	
		Size-corr. body mass	[-0.051-0.046]	0.926	-8-
Age class * Tissue + BM ratio	315	BM ratio	[-0.344-0.202]	0.609	
Age class * Tissue + migrating	315	No	[-0.050-0.048]	0.968	-8-
		Yes	[-0.165-0.040]	0.228	
Age class * Tissue + breeding	315	No	[-0.163-0.018]	0.105	
		Yes	[-0.044-0.055]	0.835	
Age class * Tissue + wintering	315	No	[-0.048-0.045]	0.960	-8-
		Yes	[-0.4060.025]	0.027	
Age class * Tissue + log([Hg] ppm dw)	315	log([Hg] ppm dw)	[-0.028-0.035]	0.809	=
Age class * Tissue + log(BMR kJ/day)	132	log(BMR kJ/day)	[-0.044-0.024]	0.560	
					-0.4 -0.2 0 0.2 0.4
					Effect of mercury on body condition

Fig. 3. Forest plot of the effect size for observed relationships between Hg concentrations and body condition in birds in correlative studies, depending on ecological, physiological and methodological factors (Fisher's ζ scores [\pm 95% confidence interval]), while accounting for tissue type, age class and their interaction. The dashed line represents a null effect size (Fisher's $\zeta = 0$). Negative effect sizes indicate lower body condition with increased Hg concentrations. The size of the squares for the mean effect size is proportional to the number of observations the effect size estimate is based on. Individual effect size and study are random factors. Model estimates are considered statistically significant if their 95% confidence interval does not cross zero. BM ratio, body mass ratio; BMR, basal metabolic rate; Body cond. index, body condition index; dw, dry mass; [Hg], Hg concentration; Other var. acc., other variables taken into account; ppm, parts per million; Sex acc., sex taken into account; Size-corr. body mass, size-corrected body mass.

living or captive bird species. However, free-ranging wintering passerines were more likely to present a negative association between Hg concentration and body condition, but further studies are necessary to confirm this trend in other taxa. We found that waders and raptors, birds in freshwater habitats and tropical/subtropical regions, and especially migrating and overwintering birds, are underrepresented in the literature on Hg contamination and body condition. Experimental studies were more likely to detect a significant effect of Hg concentrations on body condition, with 90% of estimated effect sizes being significantly positive or negative. Conversely, only 14% of correlative effect sizes for the effect of Hg contamination on body condition were significantly positive or negative. Although recent experimental studies exposed birds to environmentally realistic Hg doses, these were often in the upper range of levels encountered in the wild (Kobiela, Cristol & Swaddle, 2015; Yu *et al.*, 2016; Ma *et al.*, 2018). Therefore, the difference in sensitivity to effects between correlative and experimental studies could stem from a threshold dose that can be reached under controlled conditions, but is unlikely in the wild.

(1) Overview of experimental and correlative studies

Accumulation of energy stores can be part of the response to stressors, whereby perceived risk or unpredictable access to food can cause birds to store energy as a buffer against unpredictable environmental changes [Schultner *et al.* (2013) and

Explanatory	Sample size	Level	95% confidence interval	p–value	
log([Hg] ppm dw)	20	log([Hg] ppm dw)	[-0.253-0.225]	0.905	-#-
Dietary guild	20	Carnivore	[-1.951–0.629]	0.295	
		Herbivore	[-0.119–1.688]	0.065	
		Invertivore	[-1.169–1.410]	0.846	
Other var. acc.	20	No	[-0.336–1.644]	0.182	
		Yes	[-0.950–0.834]	0.893	
Sex acc.	20	No	[-0.360–1.192]	0.275	
		Yes	[-1.566–1.142]	0.746	
Age class	20	Adult	[-0.036–1.636]	0.060	
		Chick	[-1.313-0.532]	0.386	
Migrating		No	[-0.863–1.321]	0.665	.
		Yes	[-1.615–2.277]	0.725	
BM ratio	20	BM ratio	[-0.301–0.740]	0.389	
log(BMR kJ/day)	20	log(BMR kJ/day)	[-0.059–0.400]	0.136	-2 -15 -1 -05 0 05 1 15 2 25 Effect of mercury on body condition

Fig. 4. Forest plot of the effect size for observed relationships between Hg contamination and body condition in birds in experimental studies, depending on ecological, physiological and methodological factors (Fisher's Z scores [± 95% confidence interval]). The dashed line represents a null effect size (Fisher's Z = 0). Negative effect sizes indicate lower body condition with increased Hg exposure. The size of the squares for the mean effect size is proportional to the number of observations the effect size estimate is based on. Individual effect size and study are random factors. Model estimates are considered statistically significant if their 95% confidence interval does not cross zero. BM ratio, body mass ratio; BMR, basal metabolic rate; [Hg], Hg concentration; Other var. acc., other variables taken into account; ppm, parts per million; Sex acc., sex taken into account; ww, wet mass.

references therein]. Experimental exposure to environmentally realistic Hg levels caused an increase in energy (fat) stores and body mass in zebra finch Taeniopygia guttata (Gerson et al., 2019). Feeding rate or ingested food were not quantified, but birds were offered food ad libitum. Hence, their increase in energy stores was likely linked to an increase in food intake, as shown in mallard ducks Anas platyrhynchos exposed to Hg and fed ad libitum (Heinz, 1979). Alternatively, Hg could increase energy storage by disrupting the metabolism of carbohydrates or lipids [Seewagen (2020) and references therein]. However, in another experimental study, Hg-treated Taeniopygia guttata individuals waited longer to commence foraging and showed a significant decrease in their body mass, after exposure to predation risk (Kobiela et al., 2015). Other experimental studies showed no effect of Hg on body condition despite reduced appetite, motivation to forage, and possibly low foraging efficiency (Bouton et al., 1999; Spalding et al., 2000; Adams & Frederick, 2008). Overall, experimental studies point to disruption of feeding behaviour, and/or nutrient and energy metabolism, with positive, negative or no consequences on body condition. This suggests that metabolic effects of Hg may be weak, and thus become (statistically) detectable only at high exposure levels, and/or under specific conditions that could not be identified

by the meta-analysis. Experimental data sets were only eight in number and suffered from a slight publication bias. Therefore, we cannot exclude that further experimental work in a larger sample of individuals and species could reveal a different picture.

Previous investigations and literature reviews highlighted substantial heterogeneity in the strength and direction of the effect of Hg concentrations on body condition in wild birds: studies reported significantly negative (e.g. Ackerman et al., 2012, 2019; Fort et al., 2015; Adams et al., 2020a), positive (e.g. Kalisińska et al., 2010), or no associations (e.g. Heath & Frederick, 2005; Herring et al., 2014; Tartu et al., 2015). This heterogeneity could stem from the small statistical power of several ecotoxicological field investigations. Meta-analytical approaches can overcome this drawback and provide higher precision in the estimation of effect sizes (Koricheva et al., 2013). However, our metaanalysis confirmed the lack of a clear pattern. The correlative data set included a large number of effect sizes, with a balanced distribution of modalities for most moderators, and a lack of bias, thus suggesting that the output of this metaanalysis including both published and unpublished data is robust. Interestingly, we detected a (publication) bias towards studies that show a negative effect of Hg on body condition,

while positive effects were more likely to remain unpublished (Table S6).

Among the 15 tested moderators in correlative studies, only wintering status was identified as a driving factor of cross-study heterogeneity in results. Wintering birds were more likely to show a negative effect of Hg concentrations on body condition, suggesting a more detectable, negative effect when food is scarce and/or energetic demand for coping with unfavourable weather conditions is high. This effect was driven by two studies on several passerine species (Ackerman et al., 2012, 2019) and needs confirmation from other species. As discussed above for metabolic and behavioural effects in experimental settings, food intake and predation risk could be critical in driving effects of Hg concentration on body condition. Feeding rates, food availability and predation risk are challenging to measure in the wild, and could thus be key factors potentially confounding the Hg-body condition association in natura. In conclusion, there is a need for further studies that measure Hg-body condition associations while accounting for food intake and concurrent stressors (e.g. predation risk), especially at challenging life-cycle stages such as chick-rearing, migration, overwintering and moult.

(2) Potential confounding factors and directions for future studies

Results from our meta-analysis indicate that body condition indices are not sensitive endpoints of Hg sublethal effects in birds. In accordance with conclusions from Fuchsman et al. (2017) and Evers (2018), reproductive endpoints should be preferred to estimate Hg toxicity risk. Effects of Hg on reproductive success also have the advantage of being comparable between laboratory-based and field studies in similar taxa (Evers, 2018). The results of our meta-analysis also refute our prediction of smaller effect sizes of the Hg-body condition association in piscivorous species, because of their naturally high Hg exposure over evolutionary timescales (Scheuhammer et al., 2007; Evers, 2018). The lack of sensitivity of body condition indices to Hg effects could stem from several non-exclusive factors, some of which are inherently linked to the concept of 'body condition'. Body condition indices have been used as indicators of fat reserves, although not always explicitly so (reviewed in Labocha & Hayes, 2012). However, body mass variation can be driven largely by lean mass, not only fat mass, especially in migrating birds (Piersma, Gudmundsson & Lilliendahl, 1999; Seewagen & Guglielmo, 2011). As such, body condition indices may be poor indicators of energy stores in species with intrinsically low percentage of body lipids (Jacobs et al., 2012), and be no more informative than body mass alone (Labocha & Hayes, 2012). Here, we found no effect of body condition index type on the Hg-body condition association (Tables S2 and S4). Previous studies have shown that it is complicated to draw generalisations on which body condition index best represents body condition, but that different indices are often correlated (Labocha et al., 2014; Kraft

et al., 2019). We can speculate that if Hg had a clear impact on body condition in birds, effects would be detected irrespective of the index used, but further studies are needed to address this point specifically. In addition, body condition indices can vary substantially with season, sex and other factors, complicating comparisons among studies (Labocha & Hayes, 2012; Labocha et al., 2014). For instance, effects of Hg on body condition have been shown to depend on time of day in migrating passerines (Adams et al., 2020b), as body mass can fluctuate strongly across the day in small birds. To investigate further the potential role of energy storage and use on the relationship between Hg concentration and body condition, we tested the effect of BM ratio and BMR as moderators. The BM ratio is an indicator of maximum body condition and energy reserves (Vincze et al., 2019), while BMR represents the energy needed for basal body maintenance [in a resting, post-absorptive phase, under thermoneutral conditions (McNab, 1997; Ellis & Gabrielsen, 2001; White et al., 2007)]. Both species-specific BM ratio and BMR were poor predictors of the variation in the Hg-body condition association. However, only a third of the species included in the analysis had known BMR information, and BMR also can vary depending on other factors, such as temperature and latitude (Ellis & Gabrielsen, 2001; White et al., 2007), or the presence of other environmental contaminants such as persistent organic pollutants (Blévin et al., 2017). The influence of energy storage strategies and BMR on the association between Hg contamination and body condition needs further investigation, and likely works at the individual level, which cannot be accounted for by meta-analytical approaches.

Physiological factors could also confound the relationship between Hg concentration and body condition in birds. An example of this is the potential mismatch between the temporal integration of Hg into feathers and the timing of body condition measures (see also Section II.1). In addition, the Hg-body condition relationship could reflect mechanisms for dilution (or concentration) of Hg in tissues following body mass gain (or loss). However, this has been shown only in two studies on healthy individuals [Hg dilution in blood in growing juvenile birds (Ackerman, Eagles-Smith & Herzog, 2011); Hg concentration in blood in fasting passerines during simulated migratory fasting (Seewagen et al., 2016)], and in seabirds that died from starvation (Fort et al., 2015). Further evidence from multiple avian species is necessary to confirm whether adaptive changes in body mass and body mass composition, which are necessary to sustain energy-demanding activities such as moulting, migrating and breeding (Bech, Langseth & Gabrielsen, 1999), could drive variation in circulating Hg concentrations. To this end, we encourage the use of other non-invasive indices of body condition, such as pectoral muscle thickness (a proxy of lean mass; e.g. Sears, 1988), or body composition assessed via quantitative magnetic resonance (Seewagen et al., 2016; Ma et al., 2018), and to account for sampling time of day (Adams et al., 2020b).

Another possible factor confounding the Hg–body condition association could be selenium (Se) status. Se can play a

protective role against Hg toxicity at the biochemical level (Cuvin-Aralar & Furness, 1991; Ralston, Blackwell & Raymond, 2007; Scheuhammer et al., 2015). The formation of apparently nontoxic Hg-Se granules observed in wildlife after MeHg demethylation is considered to be the primary detoxification mechanism of MeHg, and enables long-term storage of Hg (Manceau et al., 2021). However, the mutual sequestration of Hg and Se can be detrimental. Specifically, Hg can inhibit Se-dependent enzymes (selenoenzymes), which are critical for brain health function, especially in early life (Ralston et al., 2008; Ralston, Ralston & Ravmond, 2016). Sublethal effects of Hg and the Hg-body condition association could thus be influenced by the presence and bioavailability of Se in the diet, but this is still understudied with respect to the toxic effects of Hg in avian species. Quantifying the Se:Hg molar ratio (Scheuhammer et al., 2015), and/or a risk assessment criterion that accounts for concurrent intake of MeHg and Se (Se health benefit value; Ralston et al., 2016), could improve our understanding of the sublethal effects of Hg in birds.

V CONCLUSIONS

- Our meta-analysis indicates that body condition is an unreliable endpoint of the sublethal effects of Hg in wild birds.
- (2) Associations of Hg with body condition appear to be clearer under controlled conditions and further investigations are needed.
- (3) Wintering birds were more likely to show a negative association between Hg and body condition in the wild, but further studies should confirm this in additional taxa.
- (4) We highlight a substantial knowledge gap on the metabolic effects of Hg in waders and raptors, birds in freshwater habitats and from tropical/subtropical regions, and especially in migrating and overwintering birds.
- (5) Our results indicate the need for further studies in both the laboratory and the field on the effects of Hg on feeding rates, foraging efficiency, and energy storage and use in a larger sample of individuals and species.

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VII. REFERENCES

- References identified with an asterisk (*) are cited in the online supporting information. 1. Ackerman, J. T., Eagles-Smith, C. A. & Herzog, M. P. (2011). Bird
- ACREMMAN, J. T., EAGLES-SMITH, C. A. & HERZOG, M. F. (2011). Bird mercury concentrations change rapidly as chicks age: toxicological risk is highest at hatching and fledging. *Environmental Science & Technology* 45, 5418– 5425.
- ACKERMAN, J. T., HARTMAN, C. A. & HERZOG, M. P. (2017). Maternal transfer of mercury to songbird eggs. *Environmental Pollution* 230, 463–468.
- ACKERMAN, J. T., HARTMAN, C. A. & HERZOG, M. P. (2019). Mercury contamination in resident and migrant songbirds and potential effects on body condition. *Environmental Pollution* 246, 797–810.

- 4. ACKERMAN, J. T., OVERTON, C. T., CASAZZA, M. L., TAKEKAWA, J. Y., EAGLES-SMITH, C. A., KEISTER, R. A. & HERZOG, M. P. (2012). Does mercury contamination reduce body condition of endangered California clapper rails? Environmental Pollution 162, 439-448.
- 5. ADAMS, E. M. & FREDERICK, P. C. (2008). Effects of methylmercury and spatial complexity on foraging behavior and foraging efficiency in juvenile white ibises (Eudocimus albus). Environmental Toxicology and Chemistry 27, 1708–1712.
- 6. ADAMS, E. M., SAUER, A. K., LANE, O., REGAN, K. & EVERS, D. C. (2020a). The effects of climate, habitat, and trophic position on methylmercury bioavailability for breeding New York songbirds. Ecotoxicology 29, 1843-1861.
- 7. ADAMS, E. M., WILLIAMS, K. A., OLSEN, B. J. & EVERS, D. C. (2020b). Mercury exposure in migrating songbirds: correlations with physical condition. Ecotoxicology **29**, 1240-1253.
- 8. *Albers, P. H., Koterba, M. T., Rossmann, R., Link, W. A., French, J. B., BENNETT, R. S. & BAUER, W. C. (2007). Effects of methylmercury on reproduction in American kestrels. Environmental Toxicology and Chemistry 26, 1856-1866
- 9. Albert, C., Renedo, M., Bustamante, P. & Fort, J. (2019). Using blood and feathers to investigate large-scale Hg contamination in Arctic seabirds: a review. Environmental Research 177, 108588.
- 10. Albertos, S., Berenguer, N. I., Sánchez-Virosta, P., Gómez-RAMÍREZ, P., JIMÉNEZ, P., TORRES-CHAPARRO, M. Y., VALVERDE, I., NAVAS, I., MARÍA-MOJICA, P., GARCÍA-FERNÁNDEZ, A. J. & ESPÍN, S. (2020). Mercury exposure in birds linked to marine ecosystems in the Western Mediterranean. Archives of Environmental Contamination and Toxicology 79, 435-453.
- 11. AMÉLINEAU, F., GREMILLET, D., HARDING, A. M. A., WALKUSZ, W., CHOQUET, R. & FORT, J. (2019). Arctic climate change and pollution impact little auk foraging and fitness across a decade. Scientific Reports 9, 1014.
- 12. ANDERSON, O. R. J., PHILLIPS, R. A., MCDONALD, R. A., SHORE, R. F., MCGILL, R. A. R. & BEARHOP, S. (2009). Influence of trophic position and foraging range on mercury levels within a seabird community. Marine Ecology Progress Series 375, 277-288.
- 13. *Anteau, M. J., Afton, A. D., Custer, C. M. & Custer, T. W. (2007). Relationships of cadmium, mercury, and selenium with nutrient reserves of female lesser scaup (Aythya affinis) during winter and spring migration. Environmental Toxicology and Chemistry 26, 515-520.
- 14. ATWELL, L., HOBSON, K. A. & WELCH, H. E. (1998). Biomagnification and bioaccumulation of mercury in an arctic marine food web; insights from stable nitrogen isotope analysis. Canadian Journal of Fisheries and Aquatic Sciences 55, 1114-1121.
- 15. BÅRDSEN, B.-J., HANSSEN, S. A. & BUSTNES, J. O. (2018). Multiple stressors: modeling the effect of pollution, climate, and predation on viability of a subarctic marine bird. Ecosphere 9, e02342.
- 16. BARTÓN, K. (2020). Multi-Model Inference. R package version 1.43.17. https:// CRAN.R-project.org/package=MuMIn
- 17. BASU, N., STAMLER, C. J., LOUA, K. M. & CHAN, H. M. (2005). An interspecies comparison of mercury inhibition on muscarinic acetylcholine receptor binding in the cerebral cortex and cerebellum. Toxicology and Applied Pharmacology 205, 71-76.
- 18. BECH, C., LANGSETH, I. & GABRIELSEN, G. W. (1999). Repeatability of basal metabolism in breeding female kittiwakes Rissa tridactyla. Proceedings of the Royal Society of London. Series B: Biological Sciences 266, 2161-2167.
- 19. BLÉVIN, P., TARTU, S., ELLIS, H. I., CHASTEL, O., BUSTAMANTE, P., PARENTEAU, C., HERZKE, D., ANGELIER, F. & GABRIELSEN, G. W. (2017). Contaminants and energy expenditure in an Arctic seabird: organochlorine pesticides and perfluoroalkyl substances are associated with metabolic rate in a contrasted manner. Environmental Research 157, 118–126.
- 20. BLOOM, N. S. (1992). On the chemical form of mercury in edible fish and marine invertebrate tissue. Canadian Journal of Fisheries and Aquatic Sciences 49, 1010-1017.
- 21. BOLKER, B. M., BROOKS, M. E., CLARK, C. J., GEANGE, S. W., POULSEN, J. R., STEVENS, M. H. H. & WHITE, J.-S. S. (2009). Generalized linear mixed models: a practical guide for ecology and evolution. Trends in Ecology & Evolution 24, 127-135.
- 22. BOND, A. L. & DIAMOND, A. W. (2009). Total and methyl mercury concentrations in seabird feathers and eggs. Archives of Environmental Contamination and Toxicology 56, 286-291.
- 23. BOUTON, S. N., FREDERICK, P. C., SPALDING, M. G. & MCGILL, H. (1999). Effects of chronic, low concentrations of dietary methylmercury on the behavior of juvenile great egrets. Environmental Toxicology and Chemistry 18, 1934-1939
- 24. BRAUNE, B. M., GASTON, A. J., HOBSON, K. A., GILCHRIST, H. G. & MALLORY, M. L. (2014). Changes in food web structure alter trends of mercury uptake at two seabird colonies in the Canadian Arctic. Environmental Science & Technology 48, 13246-13252.
- 25. BUSTAMANTE, P., LAHAYE, V., DURNEZ, C., CHURLAUD, C. & CAURANT, F. (2006). Total and organic Hg concentrations in cephalopods from the North

Eastern Atlantic waters: influence of geographical origin and feeding ecology. Science of The Total Environment 368, 585-596.

- 26. Carravieri, A., Burthe, S. J., de la Vega, C., Yonehara, Y., Daunt, F., NEWELL, M. A., JEFFREYS, R. M., LAWLOR, A. J., HUNT, A., SHORE, R. F., PEREIRA, M. G. & GREEN, J. A. (2020a). Interactions between environmental contaminants and gastrointestinal parasites: novel insights from an integrative approach in a marine predator. Environmental Science & Technology 54, 8938-8948.
- 27. CARRAVIERI, A., BUSTAMANTE, P., CHURLAUD, C., FROMANT, A. & CHEREL, Y. (2014a). Moulting patterns drive within-individual variations of stable isotopes and mercury in seabird body feathers: implications for monitoring of the marine environment. Marine Biology 161, 963-968.
- 28. CARRAVIERI, A., BUSTAMANTE, P., LABADIE, P., BUDZINSKI, H., CHASTEL, O. & CHEREL, Y. (2020b). Trace elements and persistent organic pollutants in chicks of 13 seabird species from Antarctica to the subtropics. Environment International 134, 105225.
- 29. CARRAVIERI, A., BUSTAMANTE, P., TARTU, S., MEILLÈRE, A., LABADIE, P., BUDZINSKI, H., PELUHET, L., BARBRAUD, C., WEIMERSKIRCH, H., CHASTEL, O. & CHEREL, Y. (2014b). Wandering albatrosses document latitudinal variations in the transfer of persistent organic pollutants and mercury to Southern Ocean predators. Environmental Science & Technology 48, 14746-14755.
- 30. CARRAVIERI, A., CHEREL, Y., BRAULT-FAVROU, M., CHURLAUD, C., Peluhet, L., Labadie, P., Budzinski, H., Chastel, O. & BUSTAMANTE, P. (2017). From Antarctica to the subtropics: contrasted geographical concentrations of selenium, mercury, and persistent organic pollutants in skua chicks (Catharacta spp.). Environmental Pollution 228, 464-473.
- 31. CARRAVIERI, A., FORT, J., TARROUX, A., CHEREL, Y., LOVE, O. P., PRIEUR, S., BRAULT-FAVROU, M., BUSTAMANTE, P. & DESCAMPS, S. (2018). Mercury exposure and short-term consequences on physiology and reproduction in Antarctic petrels. Environmental Pollution 237, 824-831.
- 32. CARRAVIERI, A., WARNER, N. A., HERZKE, D., BRAULT-FAVROU, M., TARROUX, A., FORT, J., BUSTAMANTE, P. & DESCAMPS, S. (2021). Trophic and fitness correlates of mercury and organochlorine compound residues in egg-laying Antarctic petrels. Environmental Research 193, 110518.
- 33. Chételat, J., Ackerman, J. T., Eagles-Smith, C. A. & Hebert, C. E. (2020). Methylmercury exposure in wildlife: a review of the ecological and physiological processes affecting contaminant concentrations and their interpretation. Science of The Total Environment 711, 135117.
- 34. CLARKSON, C. E., ERWIN, R. M. & RISCASSI, A. (2012). The use of novel biomarkers to determine dietary mercury accumulation in nestling waterbirds. Environmental Toxicology and Chemistry 31, 1143-1148.
- 35. Costantini, D., Bustamante, P., Brault-Favrou, M. & Dell'Omo, G. (2020). Patterns of mercury exposure and relationships with isotopes and markers of oxidative status in chicks of a Mediterranean seabird. Environmental Pollution 260, 114095.
- 36. CUVIN ARALAR, M. L. A. & FURNESS, R. W. (1991). Mercury and selenium interaction: a review. Ecotoxicology and Environmental Safety 21, 348-364.
- 37. *DE MEDEIROS COSTA, G., LÁZARO, W. L., SANPERA, C., SÁNCHEZ-FORTÚN, M., DOS SANTOS FILHO, M. & DÍEZ, S. (2021). Rhamphotheca as a useful indicator of mercury in seabirds. Science of The Total Environment 754, 141730.
- 38. DUNNING, J. B. J. JR. (2007). CRC Handbook of Avian Body Masses, Second Edition. CRC Press, Boca Raton.
- 39. EAGLES-SMITH, C. A., ACKERMAN, J. T., ADELSBACH, T. L., TAKEKAWA, J. Y., MILES, A. K. & KEISTER, R. A. (2008). Mercury correlations among six tissues for four waterbird species breeding in San Francisco Bay, California, USA. Environmental Toxicology and Chemistry 27, 2136-2153.
- 40. EAGLES-SMITH, C. A., ACKERMAN, J. T., YEE, J. & ADELSBACH, T. L. (2009). Mercury demethylation in waterbird livers: dose-response thresholds and differences among species. Environmental Toxicology and Chemistry 28, 568-577.
- 41. EAGLES-SMITH, C. A., SILBERGELD, E. K., BASU, N., BUSTAMANTE, P., DIAZ-BARRIGA, F., HOPKINS, W. A., KIDD, K. A. & NYLAND, J. F. (2018). Modulators of mercury risk to wildlife and humans in the context of rapid global change. Ambio 47, 170-197.
- 42. ECKBO, N., LE BOHEC, C., PLANAS-BIELSA, V., WARNER, N. A., SCHULL, Q., HERZKE, D., ZAHN, S., HAARR, A., GABRIELSEN, G. W. & BORGÅ, K. (2019). Individual variability in contaminants and physiological status in a resident Arctic seabird species. Environmental Pollution 249, 191-199.
- 43. EGGER, M., SMITH, G. D., SCHNEIDER, M. & MINDER, C. (1997). Bias in metaanalysis detected by a simple, graphical test. British Medical Journal 315, 629-634.
- 44. *ELBERT, R. A. & ANDERSON, D. W. (1998). Mercury levels, reproduction, and hematology in western grebes from three California Lakes, USA. Environmental Toxicology and Chemistry 17, 210-213.
- 45. Elliott, J. E., Harris, M. L., Wilson, L. K., Smith, B. D., BATCHELOR, S. P. & MAGUIRE, J. (2007). Butyltins, trace metals and morphological variables in surf scoter (Melanitta perspicillata) wintering on the south coast of British Columbia, Canada. Environmental Pollution 149, 114-124.

- EVERS, D. (2018). The effects of methylmercury on wildlife: a comprehensive review and approach for interpretation. In *The Encyclopedia of the Anthropocene* (eds D. DELLASALA and M. GOLDSTEIN), pp. 181–194. Elsevier, Oxford.
- EVERS, D. C., KEANE, S. E., BASU, N. & BUCK, D. (2016). Evaluating the effectiveness of the Minamata Convention on mercury: principles and recommendations for next steps. *Science of The Total Environment* 569–570, 888–903.
- EVERS, D. C., SAVOY, L. J., DESORBO, C. R., YATES, D. E., HANSON, W., TAYLOR, K. M., SIEGEL, L. S., COOLEY, J. H. JR., BANK, M. S. & MAJOR, A. (2008). Adverse effects from environmental mercury loads on breeding common loons. *Ecotoxicology* 17, 69–81.
- FALLACARA, D. M., HALBROOK, R. S. & FRENCH, J. B. (2011). Toxic effects of dietary methylmercury on immune function and hematology in American kestrels (*Falco sparverius*). *Environmental Toxicology and Chemistry* 30, 1320–1327.
- *FINLEY, M. T. & STENDELL, R. C. (1978). Survival and reproductive success of black ducks fed methyl mercury. *Environmental Pollution* 16, 51–64.
- FISCHER, B., TABORSKY, B. & DIECKMANN, U. (2009). Unexpected patterns of plastic energy allocation in stochastic environments. *The American Naturalist* 173, E108–E120.
- FITZGERALD, W. F., LAMBORG, C. H. & HAMMERSCHMIDT, C. R. (2007). Marine biogeochemical cycling of mercury. *Chemical Reviews* 107, 641–662.
- FORT, J., LACOUE-LABARTHE, T., NGUYEN, H. L., BOUÉ, A., SPITZ, J. & BUSTAMANTE, P. (2015). Mercury in wintering seabirds, an aggravating factor to winter wrecks? *Science of The Total Environment* 527–528, 448–454.
- 55. FORT, J., ROBERTSON, G. J., GRÉMILLET, D., TRAISNEL, G. & BUSTAMANTE, P. (2014). Spatial ecotoxicology: migratory arctic seabirds are exposed to mercury contamination while overwintering in the Northwest Atlantic. *Environmental Science & Technology* 48, 11560–11567.
- FRANCESCHINI, M. D., EVERS, D. C., KENOW, K. P., MEYER, M. W., POKRAS, M. & ROMERO, L. M. (2017). Mercury correlates with altered corticosterone but not testosterone or estradiol concentrations in common loons. *Ecotoxicology and Environmental Safety* 142, 348–354.
- FUCHSMAN, P. C., BROWN, L. E., HENNING, M. H., BOCK, M. J. & MAGAR, V. S. (2017). Toxicity reference values for methylmercury effects on avian reproduction: critical review and analysis. *Environmental Toxicology and Chemistry* 36, 294–319.
- GARCÍA, V., MORENO-OPO, R. & TINTÓ, A. (2013). Sex differentiation of Bonelli's eagle *Aquila fasciata* in Western Europe using morphometrics and plumage colour patterns. *Ardeola* 60, 261–277.
- GERSON, A. R., CRISTOL, D. A. & SEEWAGEN, C. L. (2019). Environmentally relevant methylmercury exposure reduces the metabolic scope of a model songbird. *Environmental Pollution* 246, 790–796.
- GLAHN, J. F. & MCCOY, R. B. (1995). Measurements of wintering double-crested cormorants and discriminant models of sex. *Journal of Field Ornithology* 66, 299–304.
- 61. GOUTTE, A., BARBRAUD, C., MEILLERE, A., CARRAVIERI, A., BUSTAMANTE, P., LABADIE, P., BUDZINSKI, H., DELORD, K., CHEREL, Y., WEIMERSKIRCH, H. & CHASTEL, O. (2014). Demographic consequences of heavy metals and persistent organic pollutants in a vulnerable long-lived bird, the wandering albatross. *Proceedings of the Royal Society B-Biological Sciences* 281, 20133313.
- GURNEY, K. E. B., WOOD, C. J., ALISAUSKAS, R. T., WAYLAND, M., DEVINK, J.-M. A. & SLATTERY, S. M. (2014). Identifying carry-over effects of wintering area on reproductive parameters in White-winged Scoters: an isotopic approach. *The Condor* 116, 251–264.
- HACKETT, S. J., KIMBALL, R. T., REDDY, S., BOWIE, R. C., BRAUN, E. L., BRAUN, M. J., CHOJNOWSKI, J. L., COX, W. A., HAN, K.-L. & HARSHMAN, J. (2008). A phylogenomic study of birds reveals their evolutionary history. *Science* 320, 1763–1768.
- 64. HANCOCK, J., KUSHLAN, J. A. & KAHL, M. P. (1992). Storks, Ibises and Spoonbills of the World. London: Academic Press/Harcourt Brace Jovanovich.
- HARGREAVES, A. L., WHITESIDE, D. P. & GILCHRIST, G. (2010). Concentrations of 17 elements, including mercury, and their relationship to fitness measures in arctic shorebirds and their eggs. *Science of The Total Environment* 408, 3153–3161.
- HARGREAVES, A. L., WHITESIDE, D. P. & GILCHRIST, G. (2011). Concentrations of 17 elements, including mercury, in the tissues, food and abiotic environment of Arctic shorebirds. *Science of The Total Environment* 409, 3757–3770.
- HEATH, J. A. & FREDERICK, P. C. (2005). Relationships among mercury concentrations, hormones, and nesting effort of white ibises (*Eudocimus albus*) in the Florida Everglades. *The Auk* 122, 255–267.
- *HEDDLE, C., ELLIOTT, J. E., BROWN, T. M., ENG, M. L., PERKINS, M., BASU, N. & WILLIAMS, T. D. (2020). Continuous exposure to mercury during

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embryogenesis and chick development affects later survival and reproduction of zebra finch (*Taeniopygia guttata*). *Ecotoxicology* **29**, 1117–1127.

- *HEINZ, G. (1974). Effects of low dietary levels of methyl mercury on mallard reproduction. Bulletin of Environmental Contamination and Toxicology 11, 386–392.
- *HEINZ, G. H. (1976). Methylmercury: second-year feeding effects on mallard reproduction and duckling behavior. *The Journal of Wildlife Management* 40, 82–90.
- HEINZ, G. H. (1979). Methylmercury: reproductive and behavioral effects on three generations of mallard ducks. *The Journal of Wildlife Management* 43, 394–401.
- *HEINZ, G. H., HOFFMAN, D. J., KLIMSTRA, J. D. & STEBBINS, K. R. (2010). Enhanced reproduction in mallards fed a low level of methylmercury: an apparent case of hormesis. *Environmental Toxicology and Chemistry* 29, 650–653.
- HEINZ, G. H., HOFFMAN, D. J., KLIMSTRA, J. D., STEBBINS, K. R., KONDRAD, S. L. & ERWIN, C. A. (2009). Species differences in the sensitivity of avian embryos to methylmercury. *Archives of Environmental Contamination and Toxicology* 56, 129–138.
- HELSETH, A., STERVANDER, M. & WALDENSTRÖM, J. (2005). Migration patterns, population trends and morphometrics of Ruddy Turnstones Arenaria interpres passing through Ottenby in South-Eastern Sweden. Ornis Svecica 15, 63–72.
- HERRING, G., EAGLES-SMITH, C. A., GAWLIK, D. E., BEERENS, J. M. & ACKERMAN, J. T. (2014). Physiological condition of juvenile wading birds in relation to multiple landscape stressors in the Florida everglades: effects of hydrology, prey availability, and mercury bioaccumulation. *PLoS One* 9, e106447.
- HIGGINS, J. P. & THOMPSON, S. G. (2002). Quantifying heterogeneity in a metaanalysis. *Statistics in Medicine* 21, 1539–1558.
- HIGGINS, J. P., THOMPSON, S. G., DEEKS, J. J. & ALTMAN, D. G. (2003). Measuring inconsistency in meta-analyses. *British Medical Journal* 327, 557–560.
- *HOFFMAN, D. J. & MOORE, J. M. (1979). Teratogenic effects of external egg applications of methyl mercury in the mallard, *Anas platyrhyndros. Teratology* 20, 453–461.
- *HOFFMAN, D. J., OHLENDORF, H. M., MARN, C. M. & PENDLETON, G. W. P. (1998). Association of mercury and selenium with altered glutathione metabolism and oxidative stress in diving ducks from the San Francisco bay region, USA. *Environmental Toxicology and Chemistry* 17, 167–172.
- *HOLMAN, K. L., SCHUMMER, M. L., PETRIE, S. A., CHEN, Y.-W. & BELZILE, N. (2015). Inorganic contaminants, nutrient reserves and molt intensity in autumn migrant red-necked grebes (*Podiceps grisegena*) at Georgian Bay. Archives of Environmental Contamination and Toxicology 69, 399–410.
- 81. JACKSON, A. K., EVERS, D. C., ADAMS, E. M., CRISTOL, D. A., EAGLES-SMITH, C., EDMONDS, S. T., GRAY, C. E., HOSKINS, B., LANE, O. P., SAUER, A. & TEAR, T. (2015). Songbirds as sentinels of mercury in terrestrial habitats of eastern North America. *Ecotoxicology* 24, 453–467.
- JACOBS, S. R., ELLIOTT, K., GUIGUENO, M. F., GASTON, A. J., REDMAN, P., SPEAKMAN, J. R. & WEBER, J.-M. (2012). Determining seabird body condition using nonlethal measures. *Physiological and Biochemical Zoology* 85, 85–95.
- JENNI, L. & WINKLER, R. (1989). The feather-length of small passerines: a measurement for wing-length in live birds and museum skins. *Bird Study* 36, 1–15.
- JETZ, W., THOMAS, G. H., JOY, J. B., HARTMANN, K. & MOOERS, A. O. (2012). The global diversity of birds in space and time. *Nature* 491, 444–448.
- JONES, K. C. & DE VOOGT, P. (1999). Persistent organic pollutants (POPs): state of the science. *Environmental Pollution* 100, 209–221.
- KALISIŃSKA, E., BUDIS, H., PODLASIŃSKA, J., ŁANOCHA, N. & KAVETSKA, K. M. (2010). Body condition and mercury concentration in apparently healthy goosander (*Mergus merganser*) wintering in the Odra estuary, Poland. *Ecotoxicology* **19**, 1382–1399.
- KENOW, K. P., GUTREUTER, S., HINES, R. K., MEYER, M. W., FOURNIER, F. & KARASOV, W. H. (2003). Effects of methyl mercury exposure on the growth of juvenile common loons. *Ecotoxicology* 12, 171–181.
- *KENOW, K. P., MEYER, M. W., ROSSMANN, R., GENDRON-FITZPATRICK, A. & GRAY, B. R. (2011). Effects of injected methylmercury on the hatching of common loon (*Gavia immer*) eggs. *Ecotoxicology* 20, 1684–1693.
- KOBIELA, M. E., CRISTOL, D. A. & SWADDLE, J. P. (2015). Risk-taking behaviours in zebra finches affected by mercury exposure. *Animal Behaviour* 103, 153–160.
- KOJADINOVIC, J., BUSTAMANTE, P., CHURLAUD, C., COSSON, R. P. & LE CORRE, M. (2007a). Mercury in seabird feathers: insight on dietary habits and evidence for exposure levels in the western Indian Ocean. *Science of the Total Environment* 384, 194–204.
- KOJADINOVIC, J., CORRE, M. L., COSSON, R. P. & BUSTAMANTE, P. (2007b). Trace elements in three marine birds breeding on Reunion Island (Western Indian Ocean): part 1—Factors influencing their bioaccumulation. Archives of Environmental Contamination and Toxicology 52, 418–430.
- KONISHI, M., EMLEN, S. T., RICKLEFS, R. E. & WINGFIELD, J. C. (1989). Contributions of bird studies to biology. *Science* 246, 465–472.
- KOOYMAN, G. L., SINIFF, D. B., STIRLING, I. & BENGTSON, J. L. (2004). Moult habitat, pre-and post-moult diet and post-moult travel of Ross Sea emperor penguins. *Marine Ecology Progress Series* 267, 281–290.

- KORICHEVA, J. (2003). Non-significant results in ecology: a burden or a blessing in disguise? *Oikos* 102, 397–401.
- KORICHEVA, J., GUREVITCH, J. & MENGERSEN, K. (2013). Handbook of Meta-Analysis in Ecology and Evolution. Princeton: Princeton University Press.
- KRAFT, F.-L. O. H., DRISCOLL, S. C., BUCHANAN, K. L. & CRINO, O. L. (2019). Developmental stress reduces body condition across avian life-history stages: a comparison of quantitative magnetic resonance data and condition indices. *General and Comparative Endocrinology* 272, 33–41.
- *LA SALA, L. F., PETRACCI, P. F., SMITS, J. E., BOTTÉ, S. & FURNESS, R. W. (2011). Mercury levels and health parameters in the threatened Olrog's Gull (*Larus atlanticus*) from Argentina. *Environmental Monitoring and Assessment* 181, 1–11.
- LABOCHA, M. K. & HAYES, J. P. (2012). Morphometric indices of body condition in birds: a review. *Journal of Omithology* 153, 1–22.
- LABOCHA, M. K., SCHUTZ, H. & HAYES, J. P. (2014). Which body condition index is best? *Oikos* 123, 111–119.
- 100. LERMA, M., CASTILLO-GUERRERO, J. A., RUELAS-INZUNZA, J. & FERNÁNDEZ, G. (2016). Lead, cadmium and mercury in the blood of the bluefooted booby (*Sula nebouxii*) from the coast of Sinaloa, Gulf of California, Mexico. *Marine Pollution Bulletin* **110**, 293–298.
- 101. LEWIS, S. L. & MASLIN, M. A. (2015). Defining the Anthropocene. *Nature* 519, 171–180.
- 102. LIPSEY, M. W. & WILSON, D. B. (2001). Practical Meta-Analysis. Los Angeles: SAGE Publications, Inc.
- 103. LONDOÑO, G. A., CHAPPELL, M. A., DEL CASTAÑEDA, M. R., JANKOWSKI, J. E. & ROBINSON, S. K. (2015). Basal metabolism in tropical birds: latitude, altitude, and the 'pace of life'. *Functional Ecology* **29**, 338–346.
- 104. *LUCIA, M., ANDRÉ, J.-M., BERNADET, M.-D., GONTIER, K., GÉRARD, G. & DAVAIL, S. (2008). Concentrations of metals (zinc, copper, cadmium, and mercury) in three domestic ducks in France: pekin, muscovy, and mule ducks. *Journal of Agricultural and Food Chemistry* 56, 281–288.
- 105. MA, Y., HOBSON, K. A., KARDYNAL, K. J., GUGLIELMO, C. G. & BRANFIREUN, B. A. (2021). Inferring spatial patterns of mercury exposure in migratory boreal songbirds: combining feather mercury and stable isotope (δ²H) measurements. *Science of The Total Environment* **762**, 143109.
- 106. MA, Y., PEREZ, C. R., BRANFIREUN, B. A. & GUGLIELMO, C. G. (2018). Dietary exposure to methylmercury affects flight endurance in a migratory songbird. *Environmental Pollution* 234, 894–901.
- MACCARONE, A. & BRZORAD, J. (2016). Intraspecific and intersexual variation in three species of wading birds using morphometric measurements. *Waterbirds* 39, 205–208.
- 108. MANCEAU, A., BOURDINEAUD, J.-P., OLIVEIRA, R. B., SARRAZIN, S. L. F., KRABBENHOFT, D. P., EAGLES-SMITH, C. A., ACKERMAN, J. T., STEWART, A. R., WARD-DEITRICH, C., DEL CASTILLO BUSTO, M. E., GOENAGA-INFANTE, H., WACK, A., RETEGAN, M., DETLEFS, B., GLATZEL, P., et al. (2021). Demethylation of methylmercury in bird, fish, and earthworm. *Environmental Science & Technology* **55**, 1527–1534.
- MARCOGLIESE, D. J. & PIETROCK, M. (2011). Combined effects of parasites and contaminants on animal health: parasites do matter. *Trends in Parasitology* 27, 123–130.
- 110. MARTEINSON, S. C., MARCOGLIESE, D. J. & VERREAULT, J. (2017). Multiple stressors including contaminant exposure and parasite infection predict spleen mass and energy expenditure in breeding ring-billed gulls. *Comparative Biochemistry and Physiology. Toxicology & Pharmacology: CBP* 200, 42–51.
- 111. MASON, R. P., LAPORTE, J.-M. & ANDRES, S. (2000). Factors controlling the bioaccumulation of mercury, methylmercury, arsenic, selenium, and cadmium by freshwater invertebrates and fish. *Archives of Environmental Contamination and Toxicology* 38, 283–297.
- MCKECHNE, A. E., NOAKES, M. J. & SMIT, B. (2015). Global patterns of seasonal acclimatization in avian resting metabolic rates. *Journal of Ornithology* 156, 367–376.
- MCNAB, B. K. (1997). On the utility of uniformity in the definition of basal rate of metabolism. *Physiological Zoology* 70, 718–720.
- 114. MILLS, W. F., BUSTAMANTE, P., MCGILL, R. A. R., ANDERSON, O. R. J., BEARHOP, S., CHEREL, Y., VOTTER, S. C. & PHILLIPS, R. A. (2020). Mercury exposure in an endangered seabird: long-term changes and relationships with trophic ecology and breeding success. *Proceedings of the Royal Society B: Biological Sciences* 287, 20202683.
- MøLLER, A. P. (2009). Basal metabolic rate and risk-taking behaviour in birds. *Journal of Evolutionary Biology* 22, 2420–2429.
- NOBLE, D. W. A., LAGISZ, M., O'DEA, R. E. & NAKAGAWA, S. (2017). Nonindependence and sensitivity analyses in ecological and evolutionary metaanalyses. *Molecular Ecology* 26, 2410–2425.
- 117. OBRIST, D., KIRK, J. L., ZHANG, L., SUNDERLAND, E. M., JISKRA, M. & SELIN, N. E. (2018). A review of global environmental mercury processes in

response to human and natural perturbations: changes of emissions, climate, and land use. *Ambio* **47**, 116–140.

- 118. ORTIZ-SANTALLESTRA, M. E., RESANO-MAYOR, J., HERNÁNDEZ-MATÍAS, A., RODRÍGUEZ-ESTIVAL, J., CAMARERO, P. R., MOLEÓN, M., REAL, J. & MATEO, R. (2015). Pollutant accumulation patterns in nestlings of an avian top predator: biochemical and metabolic effects. *Science of The Total Environment* 538, 692–702.
- OVERTON, C. T., CASAZZA, M. L., TAKEKAWA, J. Y. & ROHMER, T. M. (2009). Sexing California clapper rails using morphological measurements. *North American Bird Bander* 34, 58–64.
- 120. *PAROLINI, M., STURINI, M., MARASCHI, F., PROFUMO, A., COSTANZO, A., CAPRIOLI, M., RUBOLINI, D., AMBROSINI, R. & CANOVA, L. (2021). Trace elements fingerprint of feathers differs between breeding and non-breeding areas in an Afro-Palearctic migratory bird, the barn swallow (*Hinudo rustica*). *Environmental Science and Pollution Research* 28, 15828–15837.
- PEIG, J. & GREEN, A. J. (2009). New perspectives for estimating body condition from mass/length data: the scaled mass index as an alternative method. *Oikos* 118, 1883–1891.
- 122. PETERSON, S. H., ACKERMAN, J. T., TONEY, M. & HERZOG, M. P. (2019). Mercury concentrations vary within and among individual bird feathers: a critical evaluation and guidelines for feather use in mercury monitoring programs. *Environmental Toxicology and Chemistry* **38**, 1164–1187.
- PIERSMA, T., GUDMUNDSSON, G. A. & LILLIENDAHL, K. (1999). Rapid changes in the size of different functional organ and muscle groups during refueling in a long-distance migrating shorebird. *Physiological and Biochemical Zoology* 72, 405–415.
- 124. POLITO, M. J., BRASSO, R. L., TRIVELPIECE, W. Z., KARNOVSKY, N., PATTERSON, W. P. & EMSLIE, S. D. (2016). Differing foraging strategies influence mercury (Hg) exposure in an Antarctic penguin community. *Environmental Pollution* 218, 196–206.
- 125. PRAVOSUDOV, V. V. & GRUBB, T. C. (1997). Management of fat reserves and food caches in tufted titmice (*Pants tricolor*) in relation to unpredictable food supply. *Behavioral Ecology* 8, 332–339.
- 126. PROVENCHER, J. F., FORBES, M. R., HENNIN, H. L., LOVE, O. P., BRAUNE, B. M., MALLORY, M. L. & GILCHRIST, H. G. (2016a). Implications of mercury and lead concentrations on breeding physiology and phenology in an Arctic bird. *Environmental Pollution* **218**, 1014–1022.
- 127. *PROVENCHER, J. F., GILCHRIST, H. G., MALLORY, M. L., MITCHELL, G. W. & FORBES, M. R. (2016b). Direct and indirect causes of sex differences in mercury concentrations and parasitic infections in a marine bird. *Science of The Total Environment* 551–552, 506–512.
- 128. R CORE TEAM (2021). R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna.
- 129. RALSTON, N. V. C., BLACKWELL, J. L. & RAYMOND, L. J. (2007). Importance of molar ratios in selenium-dependent protection against methylmercury toxicity. *Biological Trace Element Research* 119, 255–268.
- RALSTON, N. V. C., RALSTON, C. R., BLACKWELL, J. L. & RAYMOND, L. J. (2008). Dictary and tissue selenium in relation to methylmercury toxicity. *Neuro Toxicology* 29, 802–811.
- RALSTON, N. V. C., RALSTON, C. R. & RAYMOND, L. J. (2016). Selenium health benefit values: updated criteria for mercury risk assessments. *Biological Trace Element Research* 171, 262–269.
- 132. RENEDO, M., PEDRERO, Z., AMOUROUX, D., CHEREL, Y. & BUSTAMANTE, P. (2021). Mercury isotopes of key tissues document mercury metabolic processes in seabirds. *Chemosphere* 263, 127777.
- RISING, J. D. (2010). A Guide to the Identification and Natural History of the Sparrows of the United States and Canada. London: Bloomsbury Publishing.
- 134. *ROBINSON, S. A., FORBES, M. R. & HEBERT, C. E. (2009). Parasitism, mercury contamination, and stable isotopes in fish-eating double-crested cormorants: no support for the co-ingestion hypothesis. *Canadian Journal of Zoology* 87, 740–747.
- 135. ROBINSON, S. A., FORBES, M. R., HEBERT, C. E. & SCHEUHAMMER, A. M. (2011). Evidence for sex differences in mercury dynamics in double-crested cormorants. *Environmental Science & Technology* **45**, 1213–1218.
- ROBINSON, S. A., LAJEUNESSE, M. J. & FORBES, M. R. (2012). Sex differences in mercury contamination of birds: testing multiple hypotheses with meta-analysis. *Environmental Science & Technology* 46, 7094–7101.
- 137. ROWSE, L. M., RODEWALD, A. D. & SULLIVAN, S. M. P. (2014). Pathways and consequences of contaminant flux to Acadian flycatchers (*Empidonax virescens*) in urbanizing landscapes of Ohio, USA. *Science of The Total Environment* 485–486, 461–467.
- 138. *SAGERUP, K., HELGASON, L. B., POLDER, A., STRØM, H., JOSEFSEN, T. D., SKÅRE, J. U. & GABRIELSEN, G. W. (2009). Persistent organic pollutants and mercury in dead and dying glaucous gulls (*Larus hyperboreus*) at Bjørnøya (Svalbard). *Science of The Total Environment* **407**, 6009–6016.

- SÁNCHEZ-SANTED, F., COLOMINA, M. T. & HERRERO HERNÁNDEZ, E. (2016). Organophosphate pesticide exposure and neurodegeneration. *Cortex* 74, 417–426.
- 140. SANTOS, C. S. A., BLONDEL, L., SOTILLO, A., MÜLLER, W., STIENEN, E. W. M., BOECKX, P., SOARES, A. M. V. M., MONTEIRO, M. S., LOUREIRO, S., DE NEVE, L. & LENS, L. (2017). Offspring Hg exposure relates to parental feeding strategies in a generalist bird with strong individual foraging specialization. *Science of The Total Environment* **601–602**, 1315–1323.
- 141. SANTOS, C. S. A., SOTILLO, A., GUPTA, T., DELGADO, S., MÜLLER, W., STIENEN, E. W. M., DE NEVE, L., LENS, L., SOARES, A. M. V. M., MONTEIRO, M. S. & LOUREIRO, S. (2020). Mercury uptake affects the development of *Larus fuscus* chicks. *Environmental Toxicology and Chemistry* 39, 2008–2017.
- 142. *SCHEUHAMMER, A. M. (1988). Chronic dietary toxicity of methylmercury in the zebra finch, *Poephila guttata*. Bulletin of Environmental Contamination and Toxicology 40, 123–130.
- 143. SCHEUHAMMER, A., BRAUNE, B., CHAN, H. M., FROUIN, H., KREY, A., LETCHER, R., LOSETO, L., NOËL, M., OSTERTAG, S., ROSS, P. & WAYLAND, M. (2015). Recent progress on our understanding of the biological effects of mercury in fish and wildlife in the Canadian Arctic. *Science of The Total Environment* 509–510, 91–103.
- 144. SCHEUHAMMER, A. M., LORD, S. I., WAYLAND, M., BURGESS, N. M., CHAMPOUX, L. & ELLIOTT, J. E. (2016). Major correlates of mercury in small fish and common loons (*Gavia immer*) across four large study areas in Canada. *Environmental Pollution* 210, 361–370.
- 145. SCHEUHAMMER, A. M., MEYER, M. W., SANDHEINRICH, M. B. & MURRAY, M. W. (2007). Effects of environmental methylmercury on the health of wild birds, mammals, and fish. *AMBIO: A Journal of the Human Environment* 36, 12–19.
- 146. SCHULTNER, J., KITAYSKY, A. S., WELCKER, J. & HATCH, S. (2013). Fat or lean: adjustment of endogenous energy stores to predictable and unpredictable changes in allostatic load. *Functional Ecology* 27, 45–55.
- 147. *SCHUMMER, M. L., PETRIE, S. A., BADZINSKI, S. S., CHEN, Y.-W. & BELZILE, N. (2012). Hepatic concentrations of inorganic contaminants and their relationships with nutrient reserves in autumn-migrant common loons at Lake Erie. Archives of Environmental Contamination and Toxicology 62, 704–713.
- 148. SEARS, J. (1988). Assessment of body condition in live birds; measurements of protein and fat reserves in the mute swan, *Cygnus olor. Journal of Zoology* 216, 295–308.
- 149. SEBASTIANO, M., BUSTAMANTE, P., EULAERS, I., MALARVANNAN, G., MENDEZ-FERNANDEZ, P., CHURLAUD, C., BLÉVIN, P., HAUSELMANN, A., COVACI, A., EENS, M., COSTANTINI, D. & CHASTEL, O. (2017). Trophic ecology drives contaminant concentrations within a tropical seabird community. *Environmental Pollution* 227, 183–193.
- 150. SECO, J., XAVIER, J. C., BRIERLEY, A. S., BUSTAMANTE, P., COELHO, J. P., GREGORY, S., FIELDING, S., PARDAL, M. A., PEREIRA, B., STOWASSER, G., TARLING, G. A. & PEREIRA, E. (2020). Mercury levels in Southern Ocean squid: variability over the last decade. *Chemosphere* 239, 124785.
- SEEWAGEN, C. L. (2013). Blood mercury levels and the stopover refueling performance of a long-distance migratory songbird. *Canadian Journal of Zoology* 91, 41–45.
- SEEWAGEN, C. L. (2020). The threat of global mercury pollution to bird migration: potential mechanisms and current evidence. *Ecotoxicology* 29, 1254– 1267.
- 153. SEEWAGEN, C. L., CRISTOL, D. A. & GERSON, A. R. (2016). Mobilization of mercury from lean tissues during simulated migratory fasting in a model songbird. *Scientific Reports* 6, 1–5.
- 154. SEEWAGEN, C. L. & GUGLIELMO, C. G. (2011). Quantitative magnetic resonance analysis and a morphometric predictive model reveal lean body mass changes in migrating Nearctic–Neotropical passerines. *Journal of Comparative Physiology B* 181, 413–421.
- 155. SEEWAGEN, C. L., MA, Y., MORBEY, Y. E. & GUGLIELMO, C. G. (2019). Stopover departure behavior and flight orientation of spring-migrant Yellowrumped Warblers (*Setophaga coronata*) experimentally exposed to methylmercury. *Journal of Ornithology* **160**, 617–624.
- SHIRIHAI, H., JARRETT, B., COX, J. & KIRWAN, G. M. (2002). The Complete Guide to Antarctic Wildlife: Birds and Marine Mammals of the Antarctic Continent and the Southern Ocean. London: Princeton University Press.
- 157. SOLDATINI, C., SEBASTIANO, M., ALBORES-BARAJAS, Y. V., ABDELGAWAD, H., BUSTAMANTE, P. & COSTANTINI, D. (2020). Mercury exposure in relation to foraging ecology and its impact on the oxidative status of an endangered seabird. *Science of The Total Environment* 724, 138131.
- *SPALDING, M. G., BJORK, R. D., POWELL, G. V. N. & SUNDLOF, S. F. (1994). Mercury and cause of death in great white herons. *The Journal of Wildlife Management* 58, 735–739.

- 159. SPALDING, M. G., FREDERICK, P. C., MCGILL, H. C., BOUTON, S. N. & MCDOWELL, L. R. (2000). Methylmercury accumulation in tissues and its effects on growth and appetite in captive great egrets. *Journal of Wildlife Diseases* 36, 411–422.
- 160. STERNE, J. A. C., SUTTON, A. J., IOANNIDIS, J. P. A., TERRIN, N., JONES, D. R., LAU, J., CARPENTER, J., RÜCKER, G., HARBORD, R. M., SCHMID, C. H., TETZLAFF, J., DEKKS, J. J., PETERS, J., MACASKILL, P., SCHWARZER, G., et al. (2011). Recommendations for examining and interpreting funnel plot asymmetry in meta-analyses of randomised controlled trials. *British Medical Journal* 343, d4002.
- SUKUMARAN, J. & HOLDER, M. T. (2010). DendroPy: a Python library for phylogenetic computing. *Bioinformatics* 26, 1569–1571.
- 162. SUNDERLAND, E. M., HU, X. C., DASSUNCAO, C., TOKRANOV, A. K., WAGNER, C. C. & ALLEN, J. G. (2019). A review of the pathways of human exposure to poly- and perfluoroalkyl substances (PFASs) and present understanding of health effects. *Journal of Exposure Science & Environmental Epidemiology* 29, 131–147.
- 163. *TAKEKAWA, J. Y., WAINWRIGHT-DE LA CRUZ, S. E., HOTHEM, R. L. & YEE, J. (2002). Relating body condition to inorganic contaminant concentrations of diving ducks wintering in coastal California. *Archives of Environmental Contamination and Toxicology* 42, 60–70.
- TAN, S. W., MEILLER, J. C. & MAHAFFEY, K. R. (2009). The endocrine effects of mercury in humans and wildlife. *Critical Reviews in Toxicology* 39, 228–269.
- 165. TARTU, S., ANGELIER, F., WINGFIELD, J. C., BUSTAMANTE, P., LABADIE, P., BUDZINSKI, H., WEIMERSKIRCH, H., BUSTNES, J. O. & CHASTEL, O. (2015). Corticosterone, prolactin and egg neglect behavior in relation to mercury and legacy POPs in a long-lived Antarctic bird. *Science of The Total Environment* 505, 180–188.
- 166. TARTU, S., BUSTAMANTE, P., ANGELIER, F., LENDVAI, Á. Z., MOE, B., BLÉVIN, P., BECH, C., GABRIELSEN, G. W., BUSTNES, J. O. & CHASTEL, O. (2016). Mercury exposure, stress and prolactin secretion in an Arctic seabird: an experimental study. *Functional Ecology* **30**, 596–604.
- 167. TARTU, S., BUSTAMANTE, P., GOUTTE, A., CHEREL, Y., WEIMERSKIRCH, H., BUSTNES, J. O. & CHASTEL, O. (2014). Age-related mercury contamination and relationship with luteinizing hormone in a long-lived antarctic bird. *PLoS One* 9, e103642.
- 168. TARTU, S., GOUTTE, A., BUSTAMANTE, P., ANGELIER, F., MOE, B., CLEMENT-CHASTEL, C., BECH, C., GABRIELSEN, G. W., BUSTNES, J. O. & CHASTEL, O. (2013). To breed or not to breed: endocrine response to mercury contamination by an Arctic seabird. *Biology Letters* 9, UNSP 20130317.
- 169. TCHOUNWOU, P. B., AYENSU, W. K., NINASHVILI, N. & SUTTON, D. (2003). Review: environmental exposure to mercury and its toxicopathologic implications for public health. *Environmental Toxicology* 18, 149–175.
- 170. TOBÓN, E. & OSORNO, J. L. (2006). Chick growth variability and parental provisioning in the magnificent frigatebird. *The Condor* **108**, 612–622.
- 171. UN ENVIRONMENT (2019). Global Mercury Assessment 2018. Geneva: UNEP.
- VIECHTBAUER, W. (2010). Conducting meta-analyses in R with the metafor package. *Journal of Statistical Software* 36, 1–48.
- VINCZE, O., VÁGÁSI, C. I., PAP, P. L., PALMER, C. & MØLLER, A. P. (2019). Wing morphology, flight type and migration distance predict accumulated fuel load in birds. *Journal of Experimental Biology* 222(1), jeb183517.
- VORKAMP, K., BALMER, J., HUNG, H., LETCHER, R. J. & RIGÉT, F. F. (2019). A review of chlorinated paraffin contamination in Arctic ecosystems. *Emerging Contaminants* 5, 219–231.
- 175. WADA, H., CRISTOL, D. A., MCNABB, F. M. A. & HOPKINS, W. A. (2009). Suppressed adrenocortical responses and thyroid hormone levels in birds near a mercury-contaminated river. *Environmental Science & Technology* **43**, 6031–6038.
- WALKER, C. H., SIBLY, R. M., HOPKIN, S. P. & PEAKALL, D. B. (2012). Principles of Ecotoxicology, Fourth Edition. Bosa Roca: CRC Press.
- 177. WANG, F., OUTRIDGE, P., FENG, X., MENG, B., HEIMBÜRGER, L.-E. & MASON, R. (2019). How closely do mercury trends in fish and other aquatic wildlife track those in the atmosphere? – implications for evaluating the effectiveness of the Minamata convention. *Science of the Total Environment* 674, 58–70.
- 178. *WAYLAND, M., ALISAUSKAS, R. T., KELLETT, D. K. & MEHL, K. R. (2008). Trace element concentrations in blood of nesting king eiders in the Canadian Arctic. Archives of Environmental Contamination and Toxicology 55, 683–690.
- 179. WAYLAND, M., GILCHRIST, H. G., DICKSON, D. L., BOLLINGER, T., JAMES, C., CARRENO, R. A. & KEATING, J. (2001). Trace elements in king eiders and common eiders in the Canadian Arctic. Archives of Environmental Contamination and Toxicology 41, 491–500.
- 180. WAYLAND, M., GILCHRIST, H. G., MARCHANT, T., KEATING, J. & SMITS, J. E. (2002). Immune function, stress response, and body condition in Arctic-breeding common eiders in relation to cadmium, mercury, and selenium concentrations. *Environmental Research* **90**, 47–60.

- WAYLAND, M., GILCHRIST, H. G. & NEUGEBAUER, E. (2005). Concentrations of cadmium, mercury and selenium in common eider ducks in the eastern Canadian arctic: influence of reproductive stage. *Science of The Total Environment* 351–352, 323–332.
- 182. *WAYLAND, M., HOFFMAN, D. J., MALLORY, M. L., ALISAUSKAS, R. T. & STEBBINS, K. R. (2010). Evidence of weak contaminant-related oxidative stress in glaucous gulls (*Larus hyperboreus*) from the Canadian Arctic. *Journal of Toxicology and Environmental Health, Part A* 73, 1058–1073.
- 183. WAYLAND, M., SMITS, J. J. E. G., GRANT GILCHRIST, H., MARCHANT, T. & KEATING, J. (2003). Biomarker responses in nesting, common eiders in the Canadian Arctic in relation to tissue cadmium, mercury and selenium concentrations. *Ecotoxicology* **12**, 225–237.
- 184. WEECH, S. A., SCHEUHAMMER, A. M. & ELLIOTT, J. E. (2006). Mercury exposure and reproduction in fish-eating birds breeding in the Pinchi Lake region, British Columbia, Canada. *Environmental Toxicology and Chemistry* 25, 1433–1440.
- 185. WHITE, C. R., BLACKBURN, T. M., MARTIN, G. R. & BUTLER, P. J. (2007). Basal metabolic rate of birds is associated with habitat temperature and precipitation, not primary productivity. *Proceedings of the Royal Society B: Biological Sciences* 274, 287–293.
- 186. WHITNEY, M. C. & CRISTOL, D. A. (2018). Impacts of sublethal mercury exposure on birds: a detailed review. In *Reviews of Environmental Contamination and Toxicology* (Volume **244**, ed. P. DE VOOGT), pp. 113–163. Springer International Publishing, Cham.
- 187. WILMAN, H., BELMAKER, J., SIMPSON, J., DE LA ROSA, C., RIVADENEIRA, M. M. & JETZ, W. (2014). EltonTraits 1.0: species-level foraging attributes of the world's birds and mammals. *Ecology* 95, 2027– 2027.
- WOLFE, M. F., SCHWARZBACH, S. & SULAIMAN, R. A. (1998). Effects of mercury on wildlife: a comprehensive review. *Environmental Toxicology and Chemistry* 17, 146–160.
- 189. YU, M. S., ENG, M. L., WILLIAMS, T. D., BASU, N. & ELLIOTT, J. E. (2016). Acute embryotoxic effects but no long-term reproductive effects of in ovo methylmercury exposure in zebra finches (*Taeniopygia guttata*). *Environmental Toxicology and Chemistry* 35, 1534–1540.

VIII. Supporting information

Additional supporting information may be found online in the Supporting Information section at the end of the article. **Table S1**. Relevant studies that could not be included in the meta-analysis, with reasons for exclusion.

Table S2. Test of the effect of specific body condition types on the relationship between mercury (Hg) concentration and body condition in correlative studies, while accounting for random factors 'effect ID' and 'species ID'.

Table S3. Akaike Information Criterion (AIC) selection for the best random structure of the full model for moderator analyses.

Table S4. AICc values of multifactorial models accounting for selected random factors (see Table S3) including the effect of age class, tissue and their interaction (Age class*Tissue) alone (base model) and with the addition of different moderators in the correlative data set.

Table S5. Omnibus test results of single-predictor models to quantify the drivers of variation in the effect size of the association between Hg exposure and body condition in experimental studies in birds.

Table S6. Funnel plots for published (left) and unpublished (right) correlative studies, where black dots indicate observed effect sizes and white open dots are imputed 'missing' effect sizes, based on the asymmetry of the funnel plot

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